

Package ‘cvasi’

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Type Package

Title Calibration, Validation, and Simulation of TKTD Models

Version 1.3.1

Description Eases the use of ecotoxicological effect models. Can simulate common toxicokinetic-toxicodynamic (TK/TD) models such as General Unified Threshold models of Survival (GUTS) and Lemna. It can derive effects and effect profiles (EPx) from scenarios. It supports the use of 'tidyr' workflows employing the pipe symbol. Time-consuming tasks can be parallelized.

URL <https://github.com/cvasi-tktd/cvasi>

BugReports <https://github.com/cvasi-tktd/cvasi/issues>

License GPL (>= 3)

Encoding UTF-8

LazyData true

Imports cli, rlang, stringr, dplyr, tibble, purrr, furr, tidyr, magrittr, utils, stats, methods, grid, gridExtra, ggplot2, GGally, deSolve, lubridate, units, lifecycle

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Config/testthat/edition 3

Collate 'cache_controls.R' 'class-CalibrationSet.R'
'class-ExposureSeries.R' 'class-EffectScenario.R' 'calibrate.R'
'class-ParameterSet.R' 'class-ScenarioSequence.R'
'class-Transferable.R' 'data.R' 'dose_response.R' 'effect.R'
'epx.R' 'explore_space.R' 'fx.R' 'get.R' 'globals.R' 'has.R'
'helper_functions.r' 'import_morse.R' 'import_toxswa.R' 'is.R'
'lik_profile.R' 'log.R' 'man-deb.R' 'man-lemna.R'
'man-macrophytes.R' 'solver.R' 'model-algae.R'
'model-deb_abj.R' 'model-debttox.R' 'model-deb_daphnia.R'
'model-guts_red.R' 'model-lemna_setac.R'
'model-lemna_schmitt.R' 'model-myrriophyllum.R' 'package.R'
'pll.R' 'plot.R' 'plotting.r' 'pull.R' 'sequence.R' 'set.R'
'set_bounds.R' 'set_exposure.R' 'set_forcings.R' 'set_init.R'

'set_param.R' 'set_transfer.R' 'set_window.R' 'show.R'
 'simulate.R' 'survival.R' 'utils-pipe.R'

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Author Nils Kehrein [aut, cre],

Dirk Nickisch [aut],

Peter Vermeiren [aut],

Torben Wittwer [ctb],

Johannes Witt [ctb],

Andre Gergs [ctb]

Maintainer Nils Kehrein <nils.kehrein@gmail.com>

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Algae-models	<i>Algae models</i>
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Description

Overview of supported *Algae* models

Details

- [Algae_Weber\(\)](#) by Weber *et al.* (2012)
- [Algae_TKTD\(\)](#) based on Weber *et al.* (2012), but with scaled damage
- [Algae_Simple\(\)](#) simplified growth model without additional forcing variables

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using [set_transfer\(\)](#).

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

References

Weber D, Schaeffer D, Dorgerloh M, Bruns E, Goerlitz G, Hammel K, Preuss TG and Ratte HT, 2012. Combination of a higher-tier flow-through system and population modeling to assess the effects of time-variable exposure of isoproturon on the green algae *Desmodesmus subspicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology and Chemistry*, 31, 899-908. [doi:10.1002/etc.1765](https://doi.org/10.1002/etc.1765)

EFSA Panel on Plant Protection Products and their Residues, 2018. Scientific opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms. EFSA journal 16:5377 doi:10.2903/j.efsa.2018.5377

See Also

[Lemna-models](#), [Transferable](#)

Other algae models: [Algae_Simple\(\)](#), [Algae_TKTD\(\)](#), [Algae_Weber\(\)](#)

Other models: [DEB-models](#), [GUTS-RED-models](#), [Lemna-models](#), [Macrophyte-models](#), [Myriophyllum-models](#)

Algae_Simple

Algae model with exponential growth but without additional forcings

Description

The model is a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for algae. It follows the concept of a simplified algae model described in Rendal et al. (2023). The model simulates the development of algal biomass. The growth of the algae population is simulated on the basis of growth rates, which are, in contrast to the Weber model, independent on environmental conditions which are usually optimal in laboratory effect studies. The toxicodynamic sub-model describes the effects of growth-inhibiting substances through a corresponding reduction in the photosynthesis rate on the basis of either external or internal concentrations (depending on user choice of 'scaled' parameter setting).

Usage

`Algae_Simple()`

Value

an S4 object of type [AlgaeSimple](#)

State variables

The model has two state variables:

- A, Biomass (ug fresh wt/mL, cells/mL *10⁴)
- Dw, only used if scaled = 1

Model parameters

- Growth model
 - mu_max, Maximum growth rate (d-1)
- Concentration response (Toxicodynamics)
 - EC_50, Effect concentration of 50% inhibition of growth rate (ug L-1)
 - b, slope of concentration effect curve at EC_50 (-)

- dose_response, shape of the dose response curve (0 = logit, 1 = probit)
- External concentration (Toxicokinetics)
 - kD, dominant rate constant of toxicant in aquatic environments (d-1)
 - scaled, 0 = no internal scaled damage / 1 = yes (-)

Forcings

Simplified model without additional forcings for e.g. irradiation or temperature as implemented in Algae_Weber. A constant growth over time is assumed. In case that growth is time dependent, a forcing variable (f_growth) can be set. Forcing time-series are represented by data.frame objects consisting of two columns. The first for time and the second for a scaling factor of mu_max. The input format for all forcings is a list of the data frames. If f_growth is not set, a default scaling factor of 1 is used.

Parameter boundaries

Upper and lower parameter boundaries are set by default for each parameter. This, to avoid extreme values during calibration (particularly likelihood profiling)

Simulation output

Simulation results will contain the state variables biomass (A) and scaled damage concentration (Dw).

It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`. As an example, set `nout=2` to enable reporting of external concentration (Cw) and growth scaling factor (f_growth). Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- `nout >= 1`: Cw external concentration (ug L-1)
- `nout >= 2`: f_growth growth scaling factor (-)
- `nout >= 3`: dA, biomass derivative (µg)
- `nout >= 4`: dDw, damage concentration derivative (ug L-1)

References

Weber D, Schaeffer D, Dorgerloh M, Bruns E, Goerlitz G, Hammel K, Preuss TG and Ratte HT, 2012. Combination of a higher-tier flow-through system and population modeling to assess the effects of time-variable exposure of isotroturon on the green algae *Desmodesmus subspicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology and Chemistry*, 31, 899-908. [doi:10.1002/etc.1765](https://doi.org/10.1002/etc.1765)

See Also

[Scenarios](#), [Transferable](#)

Other algae models: [Algae-models](#), [Algae_TKTD\(\)](#), [Algae_Weber\(\)](#)

Algae_TKTD	<i>Algae model with exponential growth, forcings (P, I) and scaled damage</i>
------------	---

Description

The model is a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for algae. The model simulates the development of algal biomass under laboratory and environmental conditions. The growth of the algae population is simulated on the basis of growth rates, which are dependent on environmental conditions (radiation, temperature and phosphorus). The model is a variant of the [Algae_Weber\(\)](#) model (Weber 2012) as cited in EFSA TKTD opinion (2018). This Algae model, [Algae_TKTD\(\)](#), provides an additional possibility (probit) to simulate the dose-response curve and considers a scaled internal damage instead of the external concentration.

Usage

Algae_TKTD()

Value

an S4 object of type [AlgaeTKTD](#)

State variables

The model has four state variables:

- A, Biomass (ug fresh wt/mL, cells/mL *10⁴)
- Q, Mass of phosphorous internal (ug P/ug fresh wt)
- P, Mass of phosphorous external (ug P/L)
- Dw, Damage concentration (ug/L)

Model parameters

- Growth model
 - mu_max, Maximum growth rate (d⁻¹)
 - Q_min, Minimum intracellular P (ug P/ug fresh wt)
 - Q_max, Maximum intracellular P (ug P/ug fresh wt)
 - v_max, Maximum P-uptake rate at non-limited growth (ug P/ug fresh wt/d)
 - k_s, Half-saturation constant for extracellular P (mg P/L)
 - m_max, Natural mortality rate (1/d)
 - I_opt, Optimum light intensity for growth (uE/m²/s)
 - T_opt, Optimum temperature for growth (°C)
 - T_max, Maximum temperature for growth (°C)
 - T_min, Minimum temperature for growth (°C)
- Concentration response (Toxicodynamics)

- EC₅₀, Effect concentration of 50% inhibition of growth rate (ug L⁻¹)
- b, slope of concentration effect curve at EC₅₀ (-)
- dose_resp, shape of the dose response curve (0 = logit, 1 = probit)
- External concentration (Toxicokinetics)
 - kD, dominant rate constant (d⁻¹)

Forcings

Besides exposure events (*C_w*), the *Algae* model requires two environmental properties as time-series input: Irradiance (*I*, uE/m²/s) and temperature (*T_{act}*, deg C). Forcings time-series are represented by `data.frame` objects consisting of two columns. The first for time and the second for the environmental factor in question. The input format for all forcings is a list of the data frames.

Simulation output

Simulation results will contain the state variables Biomass (*A*), mass of internal phosphorous (*Q*), mass of external phosphorous (*P*) and the damage concentration (*D_w*).

It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`. As an example, set `nout=2` to enable reporting of model derivatives *dA* and *dQ*. Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- Derivatives
 - `nout >= 1`: *dA*, biomass derivative (μg)
 - `nout >= 2`: *dQ*, internal phosphorous derivative (mg P/ug fresh wt)
 - `nout >= 3`: *dP*, external phosphorous derivative (mg P L⁻¹)
 - `nout >= 4`: *dD_w*, damage concentration derivative (ug L⁻¹)

References

Weber D, Schaeffer D, Dorgerloh M, Bruns E, Goerlitz G, Hammel K, Preuss TG and Ratte HT, 2012. Combination of a higher-tier flow-through system and population modeling to assess the effects of time-variable exposure of isoproturon on the green algae *Desmodesmus subspicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology and Chemistry*, 31, 899-908. [doi:10.1002/etc.1765](https://doi.org/10.1002/etc.1765)

See Also

[Scenarios](#), [Transferable](#)

Other algae models: [Algae-models](#), [Algae_Simple\(\)](#), [Algae_Weber\(\)](#)

Algae_Weber

*Algae model with exponential growth and forcings (I, T)***Description**

The model is a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for algae. The model simulates the development of algal biomass under laboratory and environmental conditions and was developed by Weber et al. (2012) as cited in EFSA TKTD opinion (2018). The growth of the algae population is simulated on the basis of growth rates, which are dependent on environmental conditions (radiation, temperature and phosphorus). The toxicodynamic sub-model describes the effects of growth-inhibiting substances through a corresponding reduction in the photosynthesis rate on the basis of internal concentrations. (the implementation of Weber et al. (2012) is followed where units differ with EFSA)

Usage

```
Algae_Weber()
```

Value

an S4 object of type [AlgaeWeber](#)

State variables

The model has four state variables:

- A, Biomass (ug fresh wt/mL, cells/mL *10⁴)
- Q, Mass of phosphorous internal (mg P/L, or ug P/mL)
- P, Mass of phosphorous external (mg P/L, or ug P/mL)
- C, external substance concentration (ug/L)

Model parameters

- Growth model
 - mu_max, Maximum growth rate (d⁻¹)
 - Q_min, Minimum intracellular P (ug P/ug fresh wt)
 - Q_max, Maximum intracellular P (ug P/ug fresh wt)
 - v_max, Maximum P-uptake rate at non-limited growth (ug P/ug fresh wt/d)
 - k_s, Half-saturation constant for extracellular P (mg P/L)
 - m_max, Natural mortality rate (1/d)
 - I_opt, Optimum light intensity for growth (uE/m²/s)
 - T_opt, Optimum temperature for growth (°C)
 - T_max, Maximum temperature for growth (°C)
 - T_min, Minimum temperature for growth (°C)
 - D, Dilution rate (1/d)

- R_0 , Influx concentration of P (mg P/L)
- Concentration response (Toxicodynamics)
 - EC_{50} , Effect concentration of 50% inhibition of growth rate (ug/L)
 - b , slope of concentration effect curve at EC_{50} (-)
- External concentration (Toxicokinetics)
 - k , Degradation rate of toxicant in aquatic environments (d-1)

Forcings

Besides exposure events (C_{in}), the *Algae* model requires three environmental properties as time-series input: Irradiance (I , $\mu E/m^2/s$) and temperature (T_{act} , deg C). Forcings time-series are represented by `data.frame` objects consisting of two columns. The first for time and the second for the environmental factor in question. The input format for all forcings is a list of the data frames.

Simulation output

Simulation results will contain the state variables Biomass (A), mass of internal phosphorous (Q), mass of external phosphorous (P) and the external concentration (C).

It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`. As an example, set `nout=2` to enable reporting of model derivatives dA and dQ . Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- Derivatives
 - `nout >= 1`: dA , biomass derivative (μg)
 - `nout >= 2`: dQ , internal phosphorous derivative (mg P/ug fresh wt)
 - `nout >= 3`: dP , external phosphorous derivative (mg P L-1)
 - `nout >= 4`: dC , external concentration derivative (ug L-1)

Parameter boundaries

Default values for parameter boundaries are set for all parameters by expert judgement, for calibration purposes. Values can be access from the object, and defaults overwritten.

References

- Weber D, Schaeffer D, Dorgerloh M, Bruns E, Goerlitz G, Hammel K, Preuss TG and Ratte HT, 2012. Combination of a higher-tier flow-through system and population modeling to assess the effects of time-variable exposure of isotroturon on the green algae *Desmodesmus subspicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology and Chemistry*, 31, 899-908. [doi:10.1002/etc.1765](https://doi.org/10.1002/etc.1765)
- EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, Brock T, Duquesne S, Grilli S, Hernandez-Jerez AF, Bennekou SH, Klein M, Kuhl T, Laskowski R, Machera K, Pelkonen O, Pieper S, Smith RH, Stemmer M, Sundh I,

Tiktak A, Topping CJ, Wolterink G, Cedergreen N, Charles S, Focks A, Reed M, Arena M, Ippolito A, Byers H and Teodorovic I, 2018. Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms. EFSA Journal, 16(8), 5377. doi:10.2903/j.efsa.2018.5377

See Also

[Scenarios, Transferable](#)

Other algae models: [Algae-models](#), [Algae_Simple\(\)](#), [Algae_TKTD\(\)](#)

americamysis

A DEB abj scenario of Americamysis bahia

Description

Species parameters were collected from the AddMyPet database entry on *Americamysis bahia* (Opossum shrimp). The exposure series consists of a constant exposure resulting in medium effects on length and reproduction.

Usage

```
americamysis
```

Format

An object of class `DebAbj` of length 1.

Source

https://www.bio.vu.nl/thb/deb/deblab/add_my_pet/entries_web/Americamysis_bahia/Americamysis_bahia_res.html

See Also

[DEB_abj\(\)](#)

cache_controls	<i>Cache control simulations</i>
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Description

Cache control simulations

Usage

```
cache_controls(x, windows, skipZeroExposure = FALSE, ...)
```

Arguments

x	vector of scenario objects
windows	list of window tuples
skipZeroExposure	logical, if TRUE, windows with zero exposure will not be included in calculations
...	additional parameters passed on to effect()

Value

Modified [scenario](#) objects

calibrate	<i>Fit model parameters to experimental data</i>
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Description

The function `calibrate()` performs the calibration (fitting) of model parameters to observed data. The data can originate from one or more experiments or trials. Experimental conditions, such as model parameters and exposure level, can differ between trials; fitting can be performed on all datasets at the same time.

Usage

```
calibrate(x, ...)

## S4 method for signature 'EffectScenario'
calibrate(
  x,
  par,
  data,
  endpoint = deprecated(),
  output,
```

```

    by,
    metric_fun = deprecated(),
    err_fun,
    as_tibble = deprecated(),
    catch_errors = deprecated(),
    verbose = TRUE,
    ...
)

## S4 method for signature 'CalibrationSet'
calibrate(x, par, output, err_fun, verbose = TRUE, ...)

## S4 method for signature 'list'
calibrate(
  x,
  par,
  endpoint = deprecated(),
  output,
  metric_fun = deprecated(),
  metric_total = deprecated(),
  err_fun,
  as_tibble = deprecated(),
  catch_errors = deprecated(),
  verbose = TRUE,
  ...
)

```

Arguments

x	either a single scenario or a list of caliset objects to be fitted
...	additional parameters passed on to stats::optim() and simulate()
par	named numeric vector with parameters to fit and their start values
data	<code>data.frame</code> with two or more columns with experimental data, 1st column must contain time points, the following columns may values which the scenario is fitted to.
endpoint	<i>deprecated</i> character, please use <code>output</code> instead
output	character, name of a single output column of simulate() to optimize on
by	optional character, groups and splits the experimental data into multiple distinct trials and datasets before fitting
metric_fun	<i>deprecated</i> , please use <code>err_fun</code> instead
err_fun	vectorized error function to calculate an error term that is minimized during optimization, must accept exactly four vectorized arguments, defaults to sum of squared errors
as_tibble	<i>deprecated</i> , result can no longer be returned as a tibble
catch_errors	<i>deprecated</i> , simulation errors are always caught
verbose	logical, if TRUE then debug outputs are displayed during optimization
metric_total	<i>deprecated</i>

Details

Fitting of model parameters can be performed in two ways:

1. A single [scenario](#) is fitted to a single dataset. The dataset must represent a time-series of an output variable of the model, e.g. observed biomass over time (effect data). The dataset can represent results of one or more experimental replicates under identical conditions.
2. One or more datasets of observed data are fitted each to a scenario which describes the experimental conditions during observation, such as exposure level and environmental properties. Each combination of dataset and scenario is represented by a [calibration set](#). During fitting, all *calibration sets* are evaluated and a total error term is calculated over all observed and predicted values.

Observed data:

Experimental, or effect, data must be supplied as a `data.frame` in long format with at least two columns: the first column contains numeric timestamps and the remaining columns must contain the observed quantity. The dataset must contain a column that which matches with the contents of parameter output.

As an example, the simulation result of [Lemna_Schmitt](#) model contains the output column *biomass* (BM), amongst others. To fit model parameters of said *Lemna_Schmitt* scenario based on observed biomass, the observed data must contain a column named BM which represents the observed biomass. A minimal observed dataset could look like this:

```
observed <- data.frame(time=c(0, 7, 14, 21),
                      BM=c( 12, 23, 37, 56))
```

Error function:

By default, the total sum of squared errors is used as the target function which is minimized during fitting. A custom error function can be supplied by the user: The function must accept four vectorized arguments and return a numeric of length one, i.e. the total error value which gets *minimized* by `calibrate()`.

Example of a custom error function which returns the sum of absolute errors:

```
my_absolute_error <- function(observed, predicted, weights, tags) {
  sum(abs(observed - predicted))
}
```

The arguments to the error function will contain all observed and predicted values, as well as any weights and tags that were defined by the *calibration sets*. As tags are optional, the fourth argument may be a list containing NULL values. The fourth argument can be used to pass additional information to the error function: For example, the tag may identify the study from where the data originates from and the error function could group and evaluate the data accordingly.

Value

A list of fitted parameters (as produced by `stats::optim()`) is returned.

Methods (by class)

- `calibrate(EffectScenario)`: Fit single scenario using a dataset
- `calibrate(CalibrationSet)`: Fit using a [CalibrationSet](#)
- `calibrate(list)`: Fit using a list of [caliset](#) objects

Examples

```

library(dplyr)

# Get observed biomass during control experiment by Schmitt et al. (2013)
observed <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(t, BM=obs)

# Create a scenario that represents conditions during experiment
scenario <- metsulfuron %>%
  set_param(c(k_phot_fix=TRUE, k_resp=0, Emax=1)) %>%
  set_init(c(BM=12)) %>%
  set_noexposure()

# Fit parameter 'k_phot_max' to observed biomass growth from experiment
calibrate(
  scenario,
  par=c(k_phot_max=1),
  data=observed,
  output="BM",
  method="Brent", # Brent is recommended for one-dimensional optimization
  lower=0,        # lower parameter boundary
  upper=0.5       # upper parameter boundary
) -> fit
fit$par

```

CalibrationSet

Calibration set

Description

A *calibration set* combines a [scenario](#), observed data, and an optional weighting factor into one object. The *calibration set* is used to fit model parameters to observed data using [calibrate\(\)](#).

Usage

```
caliset(scenario, data, weight = 1.0, tag = NULL)
```

Arguments

scenario	a scenario describing conditions during the experiment
data	a data.frame with observed data in long format containing two columns: the 1st column with numeric time points and 2nd column with numeric data to fit to.
weight	optional numeric weight to be applied when calculating the error term for each data point. Default value is 1.0, i.e. no weighting.
tag	optional value to identify the data, e.g. a study number

Details

A *calibration set* usually represents a single experiment or trial. Multiple experimental replicates can be combined into a single *set*, if model parameters are identical between trials. If model parameters were modified during a trial, e.g. a pump failure occurred or flow rates changed, this can be represented by using a *scenario sequence* instead of a basic [scenario](#). Please refer to [sequence\(\)](#) for details.

Weighting:

An optional weighting factor can be used to scale the error term of a whole *set* or of individual data points when fitting parameters using e.g. [calibrate\(\)](#).

The vector of weights must either be of length one or have the same length as the dataset. In the former case, the same weight will be applied to all values in the dataset. In the latter, individual weights are applied for each data point.

Value

`caliset()` returns a *calibration set* object

Examples

```
library(dplyr)

# Get observed biomass during control experiment by Schmitt et al. (2013)
observed <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(t, BM=obs)

# Create a scenario that represents conditions during experiment
scenario <- metsulfuron %>%
  set_param(c(k_phot_fix=TRUE, k_resp=0, Emax=1)) %>%
  set_init(c(BM=12)) %>%
  set_noexposure()

# Create a calibration set
cs <- caliset(scenario, observed)

# Fit parameter 'k_phot_max' to observed biomass growth from experiment
calibrate(
  cs,
  par=c(k_phot_max=1),
  output="BM",
  method="Brent", # Brent is recommended for one-dimensional optimization
  lower=0,       # lower parameter boundary
  upper=0.5     # upper parameter boundary
) -> fit
fit$par
```

DEB-models	<i>Dynamic Energy Budget (DEB) models</i>
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Description

Supported models:

- [DEB_abj](#)
- [DEBtox](#)

See Also

Other DEB models: [DEB_abj\(\)](#), [DEBtox\(\)](#)

Other models: [Algae-models](#), [GUTS-RED-models](#), [Lemna-models](#), [Macrophyte-models](#), [Myriophyllum-models](#)

DEBtox	<i>DEBtox model</i>
--------	---------------------

Description

Creates a *DEBtox* scenario as described by Jager (2020). It represents a simplified *DEBtox* model based on *DEBkiss*. In the *BYOM* application [[link](#)], this model is referred to as *DEBtox 2019*, version 4.7. It supports an optional feature of the *ERA special* model variant, which can consider a reference *Lm* parameter to compare results of multiple datasets.

Usage

`DEBtox()`

`DEB_Daphnia()`

Details

State variables:

The following list describes the names and units of the model's state variables:

- D, scaled damage ([C])
- L, body length (mm)
- R, cumulative reproduction (-)
- S, survival probability (-)

State variables D, L, and R are initialized with zero. Variable S is initialized with one (1.0). See [set_init\(\)](#) on how to set the initial state manually.

Parameters:

The following parameters are required:

- General
 - L_0 , body length at start (mm)
 - L_p , body length at puberty (mm)
 - L_m , maximum body length (mm)
 - r_B , von Bertalanffy growth rate constant (1/d)
 - R_m , maximum reproduction rate (#/d)
 - f , scaled functional response (-)
 - hb , background hazard rate (d⁻¹)
 - a , Weibull background hazard coefficient (-). Set to 1 to disable.
- Extra parameters
 - L_f , body length at half-saturation feeding (mm)
 - L_j , body length at which acceleration stops (mm)
 - T_{lag} , lag time for start development (d)
- TK/TD parameters
 - k_d , dominant rate constant (d⁻¹)
 - z_b , effect threshold energy budget ([C])
 - bb , effect strength energy-budget effects (1/[C])
 - z_s , effect threshold survival ([C])
 - bs , effect strength survival (1/[C] d)
- Other parameters (formerly globals in *BYOM*)
 - FBV , dry weight egg as fraction of structural body weight (-)
 - KRV , part. coeff. repro buffer and structure (kg/kg) (for losses with reproduction)
 - κ , approximation for kappa (for starvation response)
 - y_P , product of y_{VA} and y_{AV} (for starvation response)
 - L_{m_ref} , optional reference max length for scaling rate constants (mm). Set to zero to disable the reference length. Disabled by default.
 - len , a switch to control body length dynamics: 1 organism can shrink, 2 organism cannot shrink. Default value is 1.
 - T_{bp} , optional brood-pouch delay (d). Set to NA or zero to disable. Default value is 0.
 - MoA , mode of action switches (-). Default value is 0.
 - FB , feedback on damage dynamics switches (-). Default value is 0.

A reference L_{m_ref} is needed to properly compare different data sets, or when calibrating on more than one data set. If L_m differs, one would not want to have different rate constants at the same length.

Mode of Action:

Any combination of the following mode of actions (*MoA*) can be considered by the model:

- $MoA = 1$: assimilation/feeding
- $MoA = 2$: costs for maintenance
- $MoA = 4$: costs for growth and reproduction
- $MoA = 8$: costs for reproduction
- $MoA = 16$: hazard for reproduction

To activate more than one mode of action, simply add up the corresponding codes and set parameter MoA to the desired value. To disable all mode of actions, set parameter MoA to zero. See also [set_moa\(\)](#).

As an example, to consider effects on feeding and maintenance, set the mode of action to three (3):

```
DEBtox() %>% set_param(c(MoA=3))
```

Feedbacks:

Any combination of the following damage feedbacks can be considered by the model:

- 1: surf:vol scaling uptake rate
- 2: surf:vol scaling elimination rate
- 4: growth dilution
- 8: losses with reproduction

To activate more than one feedback, simply add up the corresponding codes. To disable all feedbacks, set the parameter to zero.

Effects:

The state variables L (body length), R (cumulative reproduction), and S (survival probability) are set as effect endpoints by default. All state variables are available as potential endpoints. The list of considered endpoints can be modified by using [set_endpoints\(\)](#).

To calculate effects, each *DEBtox* scenario is simulated twice: One simulation which considers exposure to a toxicant and one simulation without exposure, i.e. a control. See also [effect\(\)](#).

Simulation output:

The following intermediary model variables can be added to the model output on demand. Simply set the optional parameter `nout` to the required output level and pass it to [simulate\(\)](#).

- `nout >= 1`: f , actual scaled response
- `nout >= 2`: fR , actual f considering starvation
- `nout >= 3`: kd , actual kd
- `nout >= 4`: s , stress level
- `nout >= 5`: h , hazard rate
- `nout >= 6`: sA , stress factor on assimilation/feeding
- `nout >= 7`: sM , stress factor on maintenance
- `nout >= 8`: sG , stress factor on growth costs
- `nout >= 9`: sR , stress factor on reproduction costs
- `nout >= 10`: sH , stress factor on hazard to reproduction
- `nout >= 11`: xu , damage feedback factor for surf:vol scaling uptake rate
- `nout >= 12`: xe , damage feedback factor for surf:vol scaling elimination rate
- `nout >= 13`: xG , damage feedback factor for growth dilution
- `nout >= 14`: xR , damage feedback factor for losses with repro

Model history and changes:

- `cvasi v1.0.0`
 - The `DEB_Daphnia()` model implemented BYOM's *DEBtox 2019* model version 4.5
- `cvasi v1.2.0`

- The model equations were updated to conform with BYOM's *DEBtox 2019* version 4.7. This introduced a new model parameter a , the Weibull background hazard coefficient, and limited the maximum hazard rate to 99% per hour.
- The scenario constructor was renamed to `DEBtox()`.
- Additional intermediary model variables available as optional simulation output

Value

an S4 object of type `DebTox`

Functions

- `DEB_Daphnia()`: Deprecated model variant of `DEBtox()`

References

Jager T, 2020: Revisiting simplified DEBtox models for analysing ecotoxicity data. *Ecol Model* 416. doi:[10.1016/j.ecolmodel.2019.108904](https://doi.org/10.1016/j.ecolmodel.2019.108904)

Romoli et al., 2024: Environmental risk assessment with energy budget models: a comparison between two models of different complexity. *Environ Toxicol Chem* 43(2):440-449. doi:[10.1002/etc.5795](https://doi.org/10.1002/etc.5795)

See Also

Other DEB models: `DEB-models`, `DEB_abj()`

DEB_abj

DEB_abj

Description

Creates a *DEB abj* scenario. The *abj* model with type M acceleration is like model *std*, but acceleration occurs between birth and metamorphosis (V1-morph). Isomorphy is assumed before and after acceleration. Metamorphosis is before puberty and occurs at maturity E_{Hj} , which might or might not correspond with changes in morphology. The *abj* model is a one-parameter extension of model *std* ([DEB Wiki](#)).

Usage

`DEB_abj()`

Details

State variables:

The following list describes the default names and standard units of the model's state variables:

- L , structural length (cm)
- E , energy reserve (J)
- H , energy invested in maturity (J)
- R , reproduction buffer (J)
- cV , internal concentration (C)
- L_{max} , maximum structural length (cm)

All state variables are initialized with zero. See `set_init()` on how to set the initial state.

Parameters:

The following model parameters are required:

- p_M , vol-spec somatic maintenance (J/d.cm³)
- v , energy conductance (cm/d)
- k_J , maturity maint rate coefficient (1/d)
- p_{Am} , surface-area specific maximum assimilation rate (J/d.cm²)
- kap , allocation fraction to soma (-)
- E_G , spec cost for structure (J/cm³)
- f , scaled functional response (-)
- E_{Hj} , maturity at metamorphosis (J)
- E_{Hp} , maturity at puberty (J)
- kap_R , reproduction efficiency (-)
- L_b , structural length at birth (cm)
- L_j , structural length at metamorphosis (cm)
- k_e , elimination rate constant (d⁻¹)
- $c\theta$, no-effect concentration sub-lethal (C)
- cT , tolerance concentration (C)
- MoA , mode of action switch (-)

Mode of Actions:

Any combination of the following mode of actions (MoA) can be considered by the model:

- MoA = 1: effect on feeding
- MoA = 2: effect on maintenance costs
- MoA = 4: effect on overhead costs for making an egg
- MoA = 8: hazard during oogenesis
- MoA = 16: energy conductance

To activate more than one MoA, simply add up the corresponding codes. To disable all MoAs, set the parameter to zero. See also `set_mode_of_action()`.

Effects:

The state variables L (structural length) and R (reproduction buffer) are set as effect endpoints by default. All state variables are available as potential endpoints. The list of considered endpoints can be modified by using `set_endpoints()`.

To calculate effects, each *DEB* scenario is simulated twice: One simulation which considers exposure to a toxicant and one simulation without exposure, i.e. a control. See also `effect()`.

Value

an S4 object of type [DebAbj](#)

Simulation output

Simulation results will contain the state variables. It is possible to amend the output of [simulate\(\)](#) with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of [simulate\(\)](#). As an example, set `nout=2` to enable reporting of the acceleration factor (MV) and the mobilization flux (pC). Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- `nout >= 1`: MV acceleration factor (-)
- `nout >= 2`: pC mobilization flux (J/d)
- `nout >= 3`: pA assimilation flux (J/d)
- `nout >= 4`: pJ energy invested in maturity flux (J/d)

See Also

Other DEB models: [DEB-models](#), [DEBtox\(\)](#)

Examples

```
# Create an abj scenario from scratch and simulate it
DEB_abj() %>%
  set_init(c(L=0.02,E=0.1,H=0.01)) %>%
  set_param(c(p_M=3000,v=0.02,k_J=0.6,p_Am=300,kap=0.9,E_G=4000,f=1,
             E_Hj=0.05,E_Hp=0.3,kap_R=0.9,ke=1,c0=0,cT=1,L_b=0.02,
             L_j=0.04,MoA=0)) %>%
  set_exposure(no_exposure()) %>%
  set_times(0:10) %>%
  simulate()

# Print information about sample scenario 'americamysis'
americamysis

# Simulate 'americamysis' scenario
americamysis %>% simulate()
```

 dmagna

A DEBtox scenario of Daphnia magna

Description

Species and substance parameters were collected from test runs of the original [DEBtox](#) Daphnia model.

Usage

```
dmagna
```

Format

An object of class `DebTox` of length 1.

See Also

[DEBtox\(\)](#)

dose_response	<i>Calculate a dose response curve</i>
---------------	--

Description

Returns a `data.frame` with points on the dose response curve for the given effect scenario.

Usage

```
dose_response(
  scenario,
  range = c(1, 99),
  n = 20,
  strategy = c("exponential", "decadic", "vanilla"),
  verbose = FALSE,
  ...
)
```

Arguments

<code>scenario</code>	used for calculation
<code>range</code>	numeric vector specifying the required range of effect levels in percent (%), defaults to <code>c(1, 99)</code>
<code>n</code>	minimum number of points on the dose response curve
<code>strategy</code>	controls how multiplication factors are chosen, <code>vanilla</code> uses a fixed set of multiplication factors, <code>decadic</code> and <code>exponential</code> have varying step lengths.
<code>verbose</code>	logical, set to <code>TRUE</code> for additional status messages
<code>...</code>	additional arguments passed on to effect()

Details

Derives a dose response curve from a [scenario](#). The result will cover the requested range of effect levels. The tested multiplication factors can be chosen by different strategies, i.e. a `vanilla` approach using a fixed set of factors, or `decadic` and `exponential` approaches employing logarithmic and exponential factor scaling, respectively.

Value

data.frame with two columns, i.e. mf and effect

Examples

```
# basic dose response curve
minnow_sd %>% dose_response()

# modify the minimum number of points on the curve
minnow_sd %>% dose_response(n=10)

# select a subset of the effect range
minnow_sd %>% dose_response(range=c(10,20))

# use an alternative strategy for the selection of multiplication factors
minnow_sd %>% dose_response(strategy="decadic")

# provide additional output how multiplication factors were selected
minnow_sd %>% dose_response(verbose=TRUE)
```

effect	<i>Effect level</i>
--------	---------------------

Description

Derives the effect level due to toxicant exposure in the supplied scenarios. Either relative to a control scenario or derived directly from model endpoints, depending on model type. For scenarios with moving exposure windows, the maximum effect is returned.

Usage

```
effect(x, factor = 1, max_only = TRUE, ep_only = FALSE, marginal_effect, ...)
```

Arguments

x	vector of EffectScenario objects
factor	optional numeric value which scales the exposure time-series
max_only	logical, if TRUE only the maximum effect is returned, else results for all effect windows are reported
ep_only	logical, if TRUE only effect endpoints are returned as a vector
marginal_effect	numeric, if set, any effect smaller than this threshold will be reported as zero to exclude pseudo-effects originating from small numerical errors
...	additional parameters passed on to simulate()

Details

By default, only the maximum effect in all moving exposure windows will be returned. If argument `max_only=FALSE` is set, the returned table will be converted to long-format and will contain effect levels for each assessed exposure window.

Output formatting:

Start and end time of exposure windows can be disabled by setting `ep_only=TRUE`. Effect levels smaller than a certain threshold can be automatically set to zero (0.0) to avoid spurious effect levels introduced by numerical errors. Set `marginal_effect` to an adequate value less than 1%.

Computational efficiency:

Calculations can be sped up by providing a `data.frame` of pre-calculated control scenarios for each assessed time window. As control scenarios are by definition independent of any exposure multiplication factor, they can be reused for repeated calculations, e.g. to derive effect profiles or dose-response relationships.

Value

a tibble, by default containing scenarios, effect levels, and the exposure window where the maximum effect level occurred. The number of columns depends on the enabled effect endpoints and function arguments.

By default, the first column, named `scenarios`, contains the original scenario objects that were the basis of the calculation. For each effect endpoint, it will be followed by one column with the maximum effect level and two columns containing start and end time of the associated exposure window. If exposure windows are disabled, the columns will just contain the start and end time of the simulation. The effect level column will have the name of the effect endpoint, start and end time will additionally have the suffixes `.dat.start` and `.dat.end`, respectively.

 ep_x
Effect profiles (EP_x values)

Description

Derives one or more EP_x/LP_x values for the supplied effect scenarios, i.e. it calculates the multiplication factors of an exposure profile that cause x% of effect. Scenarios are processed in parallel, if possible.

Usage

```
epx(
  scenarios,
  level = c(10, 50),
  effect_tolerance = 0.001,
  factor_cutoff = NA,
  min_factor = 1e-30,
  max_factor = 1e+30,
```

```

    verbose = FALSE,
    ep_only = FALSE,
    long_format = FALSE,
    ...
  )

```

Arguments

scenarios	table or vector of <code>EffectScenario</code> objects
level	effect levels in percent (%), defaults to <code>c(10, 50)</code>
effect_tolerance	numeric, minimum absolute accuracy of effect levels
factor_cutoff	optional numeric, the search for a multiplication factor will be cut short if tried factors exceed this value; the result will report the cutoff value as the final EPx value.
min_factor	numeric, if tried factors fall below this threshold, the algorithm will halt with an error
max_factor	numeric, if tried factors exceed this threshold, the algorithm will halt with an error
verbose	logic, if TRUE then infos about model evaluations are displayed
ep_only	logical, if TRUE then only EPx values are part of the output, any contextual information such as <code>EffectScenario</code> objects are left out
long_format	logical, if TRUE then EPx values are returned as a table in long format, any contextual information will be duplicated
...	additional arguments passed on to <code>effect()</code>

Details

To estimate EPx values, a *binary search* on multiplication factors is conducted. The algorithm can achieve arbitrary precision in terms of effects. The same approach is implemented in the `morse` package in the `MFx()` function. Convergence is often achieved in less than 10 iterations per effect level and endpoint.

Internally, a knowledge base of all tried factors and resulting effect levels is kept to speed up convergence if more than one endpoint or effect level was requested. The algorithm will automatically sweep the range of multiplication factors as needed but hard cutoff values are implemented to avoid infinite loops; the algorithm will halt with an error message if tried factors are smaller than $1e-30$ or greater than $1e30$.

Numerical precision:

The precision of reported EPx values is controlled by the argument `effect_tolerance` and is given as the upper absolute error threshold of effects that is deemed acceptable. The default value of 0.001 ensures that a derived EPx will result in an effect of $x\% \pm 0.1$. Decreasing the `effect_tolerance` will result in additional model iterations and longer runtime. Setting an extremely small tolerance value may lead to a breakdown of the algorithm due to the occurrence of extremely small, quasi-random numerical errors in simulation results.

Value

The original tibble with additional columns named after the request effect levels, e.g. L.EP10. If no tibble was used as argument, then a new one is created. The first column `scenario` will contain the supplied `EffectScenario` objects.

Examples

```
minnow_sd %>% epx()
minnow_sd %>% epx(level=c(10,23,42))

# displays infos about tested multiplication factors
minnow_sd %>% epx(verbose=TRUE)

# return results as a table in wide format
minnow_sd %>% epx(long_format=TRUE)
```

epx_mtw

Calculate EPx values for a series of moving time window

Description

Calls `epx()` to calculate the EPx value (i.e. the multiplication factors of an exposure profile that cause x% of effect) for moving windows with length `window_length` that move timesteps defined by `window_interval`.

Usage

```
epx_mtw(
  x,
  level = c(10, 50),
  factor_cutoff = 1000,
  window_length = 7,
  window_interval = 1,
  ...
)
```

Arguments

<code>x</code>	a scenario
<code>level</code>	The target effect level of the effect, ie. the x of EPx.
<code>factor_cutoff</code>	above which cutoff is the EPx is not relevant
<code>window_length</code>	the length of the moving time window
<code>window_interval</code>	the interval that the moving time window moves
<code>...</code>	arguments passed to <code>epx</code>

Value

a tibble with five columns

- window.start
- window.end
- endpoint
- level
- EPx

Examples

```
metsulfuron %>%  
  set_window(length=7, interval=1) %>%  
  ep_x_mtw()
```

explore_space

Explore parameter space

Description

The function is aimed at getting an idea of how the parameter space of a model behaves, so that parameter identifiability problems and correlations between parameters can be explored. Therefore, the function samples a large number of parameter sets by randomly drawing from each parameter's 95% confidence interval (generated by [lik_profile\(\)](#)). It then checks how many of the parameter sets are within acceptable limits by comparing the likelihood ratio of a parameter set vs. the original parameter set against a chi-square distribution as degrees of freedom (df) the total number of profile parameters (outer rim) or one df (inner rim). If needed, the function resamples until at least `nr_accept` parameters sets are within the inner rim

Usage

```
explore_space(  
  x,  
  par,  
  res,  
  output,  
  sample_size = 1000,  
  max_runs = 30,  
  nr_accept = 100,  
  sample_factor = 1.2  
)
```

Arguments

x	a list of <code>caliset</code> objects
par	best fit parameters from joined calibration
res	output of 'lik_profile()' function
output	character vector, name of output column of <code>simulate()</code> that is used in calibration
sample_size	number of samples to draw from each parameter interval
max_runs	max number of times to redraw samples (within a smaller space), and repeat the process
nr_accept	threshold for number of points sampled within the inner circle
sample_factor	multiplication factor for sampling (95% interval * sample factor)

Value

a list containing a plot to explore the parameter space, and the data.frame supporting it

Examples

```
library(dplyr)
# Example with Lemna model - physiological params
# Before applying the function, a model needs to be calibrated and its parameters profiled
# Inputs for likelihood profiling

# exposure - control run
exp <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(time=t, conc)

# observations - control run
obs <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(t, BM=obs)

# parameters after calibration
params <- c(
  k_phot_max = 5.663571,
  k_resp = 1.938689,
  Topt = 26.7
)

# set parameter boundaries (if different from defaults)
bounds <- list(
  k_resp = list(0, 10),
  k_phot_max = list(0, 30),
  Topt = list(20, 30)
)

# update metsulfuron
myscenario <- metsulfuron %>%
```

```

set_init(c(BM = 5, E = 1, M_int = 0)) %>%
set_param(list(
  k_0 = 5E-5,
  a_k = 0.25,
  BM50 = 17600,
  mass_per_fronnd = 0.1
)) %>%
set_exposure(exp) %>%
set_param(params) %>%
set_bounds(bounds)

# Likelihood profiling
res <- lik_profile(
  x = myscenario,
  data = obs,
  output = "BM",
  par = params,
  refit = FALSE,
  type = "fine",
  method = "Brent"
)
# plot
plot_lik_profile(res)

# parameter space explorer
set.seed(1) # for reproducibility
res_space <- explore_space(
  x = list(caliset(myscenario, obs)),
  par = params,
  res = res,
  output = "BM",
  sample_size = 1000,
  max_runs = 20,
  nr_accept = 100)

plot_param_space(res_space)

```

ExposureSeries

Exposure time-series

Description

Creates an object that encapsulates an exposure time-series with its metadata, such as formatted datetime strings and file name where the series was loaded from. `no_exposure()` is shorthand to create a time-series of constant zero exposure.

Usage

```
ExposureSeries(series, dates, file, meta, context)
```

Arguments

series	data.frame with two columns containing a time-series
dates	vector, optional original list of time stamps
file	character, optional file name where data originates from
meta	list, optional metadata
context	list optional contextual metadata such as project ids

Value

an S4 object of type [ExposureSeries](#)

Slots

dates	original time points of time-series, e.g. time stamps of the form 2000-01-01 12:00
file	character, file name where data originates from, may be empty
meta	list, contains metadata
context	list, contains contextual metadata, such as project ids
series	data.frame containing the actual time-series

See Also

[no_exposure\(\)](#)

focusd1

A Lemna_SETAC scenario with variable environment

Description

A mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for the aquatic macrophytes *Lemna* spp. as published by Klein *et al.* (2021).

Usage

```
focusd1
```

Format

An object of class `LemnaSetac` of length 1.

Details

The scenario will simulate a period of 365 days, a start population of 80 g/m² dry weight, variable environmental conditions, and a complex, time-varying exposure pattern.

The scenario setup was published by Hommen *et al.* (2015). Exposure pattern and substance specific parameters are of exemplary character and represent the herbicide *metsulfuron-methyl*. The parameters were derived by Schmitt *et al.* (2013) based on literature data.

References

Hommen U., Schmitt W., Heine S., Brock Theo CM., Duquesne S., Manson P., Meregalli G., Ochoa-Acuña H., van Vliet P., Arts G., 2015: How TK-TD and Population Models for Aquatic Macrophytes Could Support the Risk Assessment for Plant Protection Products. *Integr Environ Assess Manag* 12(1), pp. 82-95. doi:10.1002/ieam.1715

Klein J., Cedergreen N., Heine S., Reichenberger S., Rendal C., Schmitt W., Hommen U., 2021: Refined description of the *Lemna* TKTD growth model based on *Schmitt et al.* (2013) - equation system and default parameters. Report of the working group *Lemna* of the SETAC Europe Interest Group Effect Modeling. Version 1, uploaded on 22. Sept. 2021. <https://www.setac.org/group/effect-modeling.html>

Schmitt W., Bruns E., Dollinger M., Sowig P., 2013: Mechanistic TK/TD-model simulating the effect of growth inhibitors on *Lemna* populations. *Ecol Model* 255, pp. 1-10. doi:10.1016/j.ecolmodel.2013.01.017

See Also

[Lemna-models](#)

Examples

```
# Simulate the example scenario
focusd1 %>% simulate()
```

fx

Generic to calculate effects for a particular scenario

Description

Generic to calculate effects for a particular scenario

Usage

```
fx(scenario, ...)

## S4 method for signature 'ANY'
fx(scenario, ...)

## S4 method for signature 'Algae'
fx(scenario, ...)

## S4 method for signature 'GutsRedSd'
fx(scenario, ...)

## S4 method for signature 'GutsRedIt'
fx(scenario, ...)
```



```
## S4 method for signature 'Lemna'
fx(scenario, ...)

## S4 method for signature 'Myriophyllum'
fx(scenario, ...)
```

Arguments

scenario [scenario](#) object
 ... additional parameters

Value

numeric named vector

Methods (by class)

- fx(ANY): Use state variables at end of simulation
- fx(Algae): Effect at end of simulation of [Algae-models](#)
- fx(GutsRedSd): Survival and lethality in [GUTS-RED-models](#)
- fx(GutsRedIt): Survival and lethality in [GUTS-RED-models](#)
- fx(Lemna): Effect at end of simulation of [Lemna-models](#)
- fx(Myriophyllum): Effect at end of simulation of [Myriophyllum-models](#)

get_model	<i>Get model name</i>
-----------	-----------------------

Description

Returns the unique model name that is associated with a scenario, e.g. GUTS-RED-IT. The function supports vectorized arguments.

Usage

```
get_model(x)
```

Arguments

x (vector of) [scenarios](#) or [parameter_set](#) objects

Value

vector of character

Examples

```
# returns `GUTS-RED-IT`
get_model(minnow_it)
```

get_tag	<i>Get scenario tag</i>
---------	-------------------------

Description

Returns the user-defined, custom tag of a scenario, if available. Tags can be helpful to quickly distinguish scenarios of the same model type. The function supports vectorized inputs.

Usage

```
get_tag(x)
```

Arguments

x (vector of) [scenarios](#) or [parameter_set](#) objects

Value

vector of character

See Also

[set_tag\(\)](#)

Examples

```
# returns `fathead minnow`
get_tag(minnow_it)

# update or set a tag
myscenario <- minnow_it %>% set_tag("My Custom Tag")
# returns `My Custom Tag`
get_tag(myscenario)
```

GUTS-RED-models

GUTS-RED models

Description

Reduced *General Unified Threshold models of Survival* (GUTS) with stochastic death (*SD*) and individual tolerance (*IT*)

Details

The TKTD models *GUTS-RED-SD* and *GUTS-RED-IT* were described by EFSA (2018). GUTS-RED models assume a one-compartment model which directly links external concentration to the scaled damage. The scaled damage is given in units of concentration, equal to the units of measurement in the external medium, e.g. ug/L. The damage dynamics is connected to an individual hazard state variable, resulting in simulated mortality when an internal damage threshold is exceeded. The death mechanisms stochastic death (*SD*) and individual threshold (*IT*) are extreme cases of the *GUTS* theory.

For *SD* models, the threshold parameter for lethal effects is fixed and identical for all individuals of a group, meaning that the variance of the threshold values is zero. Hence, the killing rate relates the probability of a mortality event in proportion to the scaled damage. For *IT* models, the thresholds for effects are distributed among individuals of a group. Mortality of an individual follows immediately once the individual's tolerance is exceeded. Meaning in model terms that the killing rate is set to infinity (EFSA 2018).

State variables

The following list describes the default names and standard units of *GUTS-RED* state variables:

- D, scaled damage (conc)
- H, cumulative hazard (-)

The state variables are initialized with zero by default.

SD model parameters

- kd, dominant rate constant (time⁻¹)
- hb, background hazard rate (time⁻¹)
- z, threshold for effects (conc)
- kk, killing rate constant (time⁻¹)

IT model parameters

- kd, dominant rate constant (time⁻¹)
- hb, background hazard rate (time⁻¹)
- alpha, median of thresholds (conc)
- beta, shape parameter (-)

Effects

The effect endpoint L (lethality) is available for *GUTS-RED* models. A value of zero (0.0) denotes *no effect* on organism survival. A value of one (1.0) denotes a lethality rate of 100%, i.e. no survivors.

The survival probability S is available in the return value of `simulate()`.

References

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

Other GUTS-RED models: [GUTS_RED_IT\(\)](#), [GUTS_RED_SD\(\)](#)

Other models: [Algae-models](#), [DEB-models](#), [Lemna-models](#), [Macrophyte-models](#), [Myriophyllum-models](#)

GUTS_RED_IT

GUTS-RED-IT scenario

Description

Reduced *General Unified Threshold models of Survival* (GUTS) with individual tolerance (*IT*).

Usage

GUTS_RED_IT(param, init)

Arguments

param	optional named list or vector with model parameters
init	optional named numeric vector to use as initial state

Value

an S4 object of type [GutsRedIt](#)

Simulation output

The return value of [simulate\(\)](#) will contain values for the state variables, as well as an additional column S which represents the survival probability for each time point. S is calculated as described in EFSA (2018) as $S = (I - F(t))$. The background hazard rate hb is already considered in state variable H and therefore does not occur as an additional term to derive S.

State variables

The following list describes the default names and standard units of *GUTS-RED* state variables:

- D, scaled damage (conc)
- H, cumulative hazard (-)

The state variables are initialized with zero by default.

IT model parameters

- kd, dominant rate constant (time⁻¹)
- hb, background hazard rate (time⁻¹)
- alpha, median of thresholds (conc)
- beta, shape parameter (-)

Effects

The effect endpoint L (lethality) is available for *GUTS-RED* models. A value of zero (0.0) denotes *no effect* on organism survival. A value of one (1.0) denotes a lethality rate of 100%, i.e. no survivors.

The survival probability S is available in the return value of `simulate()`.

References

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

Other GUTS-RED models: [GUTS-RED-models](#), [GUTS_RED_SD\(\)](#)

GUTS_RED_SD

GUTS-RED-SD scenario

Description

Reduced *General Unified Threshold models of Survival* (GUTS) with stochastic death (SD).

Usage

```
GUTS_RED_SD(param, init)
```

Arguments

param	optional named list or vector with model parameters
init	optional named numeric vector to use as initial state

Value

an S4 object of type [GutsRedSd](#)

Simulation output

The return value of `simulate()` will contain values for the state variables, as well as an additional column *S* which represents the survival probability for each time point. *S* is calculated as described in EFSA (2018) as $S = \exp(-H)$. The background hazard rate *hb* is already considered in state variable *H* and therefore does not occur as an additional term to derive *S*.

State variables

The following list describes the default names and standard units of *GUTS-RED* state variables:

- *D*, scaled damage (conc)
- *H*, cumulative hazard (-)

The state variables are initialized with zero by default.

SD model parameters

- *kd*, dominant rate constant (time⁻¹)
- *hb*, background hazard rate (time⁻¹)
- *z*, threshold for effects (conc)
- *kk*, killing rate constant (time⁻¹)

Effects

The effect endpoint *L* (lethality) is available for *GUTS-RED* models. A value of zero (0.0) denotes *no effect* on organism survival. A value of one (1.0) denotes a lethality rate of 100%, i.e. no survivors.

The survival probability *S* is available in the return value of `simulate()`.

References

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

Other GUTS-RED models: `GUTS-RED-models`, `GUTS_RED_IT()`

import_morse	<i>Import morse model parameters</i>
--------------	--------------------------------------

Description

Loads GUTS model parameters which were fitted by the morse package.

Usage

```
import_morse(
  fit,
  find_sd = TRUE,
  find_it = TRUE,
  reset_hb = TRUE,
  params = c("estim", "all"),
  mcmc_size,
  find.SD = deprecated(),
  find.IT = deprecated(),
  reset.hb = deprecated(),
  mcmc.size = deprecated(),
  file = deprecated()
)

morse(...)
```

Arguments

fit	Either a string with a file path to an <i>.Rdata</i> or <i>.RDS</i> file containing a <i>morse</i> fit, or a <i>morse</i> fit object itself
find_sd	a logical value. If TRUE, it will try to find fitted parameters of a <i>GUTS-RED-SD</i> model
find_it	a logical value. If TRUE, it will try to find fitted parameters of a <i>GUTS-RED-IT</i> model
reset_hb	a logical value. If TRUE, the background hazard rate hb is set to zero
params	character, if set to "estim" then only the best-fit parameters are imported, else all parameter sets in the MCM chains are returned
mcmc_size	optional integer, sets the maximum number of imported parameter sets per MCMC. By default, all MSMS parameter samples are imported.
find.SD	<i>deprecated</i> , alias for parameter find_sd
find.IT	<i>deprecated</i> , alias for parameter find_it
reset.hb	<i>deprecated</i> , alias for parameter rest_hb
mcmc.size	<i>deprecated</i> , alias for parameter mcmc_size
file	<i>deprecated</i> , alias for parameter fit
...	Arguments passed on to import_morse()

Value

list of [parameter_set](#) objects

Functions

- `morse()`: deprecated alias

Examples

```
# import all parameter fits
try(import_morse("path/to/morse_fit.RData"))

# import parameters for a specific model
try(import_morse("path/to/morse_fit.RData", find_it=TRUE, find_sd=FALSE))

# modify model objects
try(models %>% set_param(import_morse("path/to/morse_fit.RData")))
```

import_swash

SWASH project exposure profile import

Description

Read all TOXSWA files within a SWASH project directory.

Usage

```
import_swash(swash_dir, ...)
```

Arguments

<code>swash_dir</code>	path to the SWASH project directory
<code>...</code>	arguments passed on to import_toxswa()

Value

a list of imported exposure series, see [import_toxswa\(\)](#) for details

import_toxswa	<i>Import TOXSWA exposure series</i>
---------------	--------------------------------------

Description

Read one or more *TOXSWA* exposure series from *TOXSWA*'s .out files. By default, the concentration dissolved in water (*ConLiqWatLay*) at the end of the simulated waterbody (i.e. at the maximum of the *x* dimension) is returned. The unit of the time scale as well as of the imported model output variable can be scaled as needed.

Usage

```
import_toxswa(
  files,
  alias = NA,
  output_var = "ConLiqWatLay",
  output_unit = "ug/L",
  time_unit = "days",
  substance = NULL,
  split = TRUE
)
```

Arguments

<code>files</code>	vector of strings with absolute or relative paths to files
<code>alias</code>	optional vector with strings, will be used as an alias to identify a <i>TOXSWA</i> series instead of its filename
<code>output_var</code>	character, single output variable from <i>TOXSWA</i> that is imported, defaults to <i>ConLiqWatLay</i>
<code>output_unit</code>	character, target unit of the imported output variable, defaults to <i>ug/L</i> , syntax must be compatible with <code>units::units()</code>
<code>time_unit</code>	character, target unit of the imported time scale, defaults to <i>days</i> , syntax must be compatible with <code>units::units()</code>
<code>substance</code>	optional vector of characters, if set, only the substance codes defined in this vector are imported
<code>split</code>	logical, if TRUE then one series will be returned for each substance found in the <i>TOXSWA</i> files, else all substances per file will be in one <i>data.frame</i> . Defaults to TRUE

Details

The numerical time scale is shifted to always start at time zero (0.0). Numerical columns of the returned *data.frame* objects will be of type `units::units`. Please be aware that the use of `units` objects may not be supported by all functions in this package. However, `set_times()` and `set_exposure()` can handle `units` objects safely.

Incomplete list of alternative *TOXSWA* v5.5.3 output variables:

- *ConLiqWatLay*: Concentration dissolved in water (g/m3)
- *ConLiqSed*: Concentration in pore water sediment (g/m3)
- *ConSysWatLay*: Total concentration in water (g/m3)
- *CntSorSusSol*: Content sorbed to suspended solids (g/kg)
- *CntSorSed*: Content sorbed to sediment (g/kg)

Value

list of *data.frame* objects with exposure series. Each *data.frame* has at least three columns:

- *time*: numerical time scale, always starts at zero
- *timestamp*: time as datetime objects such as POSIXct
- one or more additional columns for each imported substance

is_DEB

Test if argument is a DEB model

Description

Test if argument is a DEB model

Usage

is_DEB(x)

Arguments

x vector of EffectScenario objects

Value

vector of logical values

is_GUTS	<i>Test if argument is a GUTS model</i>
---------	---

Description

Test if argument is a GUTS model

Usage

```
is_GUTS(x)
```

```
is_GUTS_IT(x)
```

```
is_GUTS_SD(x)
```

Arguments

x vector of EffectScenario objects

Value

vector of logical values

Functions

- `is_GUTS_IT()`: Test if argument is a GUTS-IT model
- `is_GUTS_SD()`: Test if argument is a GUTS-IT model

Examples

```
# returns `TRUE`  
is_GUTS(minnow_it)  
is_GUTS(GUTS_RED_IT())  
  
# returns `c(TRUE,TRUE,TRUE)`  
is_GUTS(c(minnow_it, minnow_it, minnow_it))  
  
# returns `FALSE`  
is_GUTS_SD(minnow_it)
```

is_Lemna *Test if argument is a Lemna model*

Description

Also returns TRUE for LemnaThreshold models

Usage

```
is_Lemna(x)
```

Arguments

x vector of [scenarios](#) objects

Value

vector of logical values

See Also

[is_LemnaThreshold\(\)](#)

is_LemnaThreshold *Test if argument is a LemnaThreshold model*

Description

Test if argument is a LemnaThreshold model

Usage

```
is_LemnaThreshold(x)
```

Arguments

x vector of [scenarios](#) objects

Value

vector of logical values

See Also

[is_Lemna\(\)](#)

is_scenario	<i>Test if argument is an effect scenario</i>
-------------	---

Description

Supports vectorized arguments.

Usage

```
is_scenario(x)
```

Arguments

x Some value or object

Value

vector of logical values

Examples

```
# returns `TRUE`  
is_scenario(minnow_it)  
  
# returns `FALSE`  
is_scenario(list())
```

Lemna-models	<i>Lemna models</i>
--------------	---------------------

Description

Overview of supported *Lemna* models

Details

- [Lemna_Schmitt\(\)](#) by Schmitt *et al.* (2013)
- [Lemna_SETAC\(\)](#) by Klein *et al.* (2021)

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

See Also

[Macrophyte-models](#)

Other Lemna models: [Lemna_SETAC\(\)](#), [Lemna_Schmitt\(\)](#)

Other models: [Algae-models](#), [DEB-models](#), [GUTS-RED-models](#), [Macrophyte-models](#), [Myriophyllum-models](#)

Lemna_Schmitt

Lemna model (Schmitt et al. 2013)

Description

The model is a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for the aquatic macrophytes *Lemna spp.* The model simulates the development of *Lemna* biomass under laboratory and environmental conditions and was developed by Schmitt *et al.* (2013). Growth of the *Lemna* population is simulated on basis of photosynthesis and respiration rates which are functions of environmental conditions. The toxicodynamic sub-model describes the effects of growth-inhibiting substances by a respective reduction in the photosynthesis rate based on internal concentrations. This is the historical version of the Lemna model. For current uses, we recommend the Lemna (SETAC) model, which is a more recent version of the Schmitt model.

Usage

```
Lemna_Schmitt(param, init)
```

```
Lemna_SchmittThold(param, init)
```

Arguments

param	optional named list or vector of model parameters
init	optional named numeric vector of initial state values

Details

Constructors to ease creation of scenarios based on the *Lemna* model by Schmitt *et al.* (2013). A variant of this *Lemna* model, `Lemna_SchmittThold()`, provides an additional cumulative exposure threshold parameter. The Lemna biomass stops growing if the integral of exposure over time exceeds the threshold. The integral of exposure is internally accounted for by an additional state variable AUC (Area Under Curve).

Value

an S4 object of type `LemnaSchmitt`

Functions

- `Lemna_SchmittThold()`: model variant with cumulative exposure threshold

State variables

The following list describes the default names and standard units of the model's state variables:

- BM, g_dw/m², dry weight biomass per square meter
- E, -, effect [0,1]
- M_int, ug, internal toxicant mass
- AUC, ug/L, cumulative exposure (**only** for LemnaThreshold model)

Biomass (BM) and internal toxicant mass (M_int) are initialized to zero by default. See `set_init()` on how to set the initial states.

Model parameters

The following model parameters are required:

- Fate and biomass
 - `k_phot_fix`, logical, TRUE then `k_phot_max` is not changed by environmental factors, else FALSE
 - `k_phot_max`, 1/d, maximum photosynthesis rate
 - `k_resp`, 1/d, respiration rate
 - `k_loss`, 1/d, rate of loss (e.g. flow rate)
 - `mass_per_fron`, g_dw/frond, dry weight per frond
 - `BMw2BMd`, g_fw/g_dw, Fresh weight/dry weight
- Effect
 - `E_max`, -, maximum effect [0,1]
 - `EC50`, ug/L, midpoint of effect curve
 - `b`, -, slope of effect curve
- Toxicokinetics
 - `P_up`, cm/d, Permeability for uptake
 - `AperBM`, cm²/g_dw, $A_{leaf} / d_{leaf} = 1/d_{leaf}$ (for circular disc, d=0.05 cm)

- K_{bm} , -, Biomass(fw) : water partition coefficient
- P_{Temp} , logical, TRUE to enable temperature dependence of cuticle permeability, else FALSE
- $MolWeight$, g/mol, Molmass of molecule (determines $Q_{10_permeability}$)
- Temperature dependence
 - T_{min} , deg C, minimum temperature for growth
 - T_{max} , deg C, maximum temperature for growth
 - T_{opt} , deg C, optimal temperature for growth
 - t_{ref} , deg C, reference temperature for respiration rate
 - Q_{10} , -, temperature dependence factor for respiration rate
- Light dependence
 - k_0 , 1/d, light dependence: intercept of linear part
 - a_k , (1/d)/(kJ/m².d), light dependence: slope of linear part
- Phosphorus dependence (Hill like dep.)
 - C_P , mg/L, phosphorus concentration in water
 - CP_{50} , mg/L, phosphorus conc. where growth rate is halved
 - a_p , -, Hill coefficient
 - K_{iP} , mg/L, p-inhibition constant for very high p-conc.
- Nitrogen dependence (Hill like dep.)
 - C_N , mg/L, nitrogen concentration in water
 - CN_{50} , mg/L, n-conc. where growth rate is halved
 - a_N , -, Hill coefficient
 - K_{iN} , mg/L, n-inhibition constant for very high p-conc.
- Density dependence
 - BM_{50} , g_{dw}/m², cut off BM

The Lemna_SchmittThold model requires the following additional parameter:

- $threshold$, ug/L, cumulative exposure threshold

Forcings

Besides exposure, the Lemna model requires two environmental properties as time-series input: global radiation (rad, kJ/m².d) and temperature (temp, deg C). Forcings time-series are represented by `data.frame` objects consisting of two columns. The first for time and the second for the environmental factor in question.

Entries of the `data.frame` need to be ordered chronologically. A time-series can consist of only a single row; in this case it will represent constant environmental conditions. See [scenarios](#) for more details.

Effects

Supported effect endpoints include *BM* (biomass) and *r* (average growth rate during simulation). The effect on biomass is calculated from the last state of a simulation. Be aware that endpoint *r* is incompatible with frond transfers.

Parameter boundaries

Default values for parameter boundaries are set for all parameters by expert judgement, for calibration purposes. Values can be accessed from the object, and defaults overwritten.

Simulation output

Simulation results will contain two additional columns besides state variables:

- C_int, ug/L, internal concentration of toxicant
- FrondNo, -, number of fronds

It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`, see examples below. `nout=1` enables reporting of internal concentration (C_int), `nout=14` enables all additional outputs, and `nout=0` will disable additional outputs.

The available output levels are as follows:

- `nout >= 1`: C_int, internal concentration (ug/L)
- `nout >= 2`: FrondNo, number of fronds (-)
- `nout >= 3`: C_int_u, unbound internal concentration (ug/l)
- Growth and TK/TD
 - `nout >= 4`: BM_fresh, fresh weight biomass (g_fw/m2)
 - `nout >= 5`: k_photo_eff, current photosynthesis rate (1/d)
 - `nout >= 6`: k_resp_eff, current respiration rate (1/d)
 - `nout >= 7`: f_Eff, toxic effect factor (-)
 - `nout >= 8`: P_up_eff, current permeability for uptake (cm/d)
- Environmental variables
 - `nout >= 9`: actConc, current toxicant concentration in surrounding medium (ug/L)
 - `nout >= 10`: actTemp, current environmental temperature (deg C)
 - `nout >= 11`: actRad, current environmental radiation (kJ/m2.d)
- Derivatives
 - `nout >= 12`: d BM/dt, current change in state variable BM
 - `nout >= 13`: d E/dt, current change in effect
 - `nout >= 14`: d M_int/dt, current change in internal toxicant mass

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g.

reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

References

Schmitt W., Bruns E., Dollinger M., and Sowig P., 2013: *Mechanistic TK/TD-model simulating the effect of growth inhibitors on Lemna populations*. Ecol Model 255, pp. 1-10. doi:10.1016/j.ecolmodel.2013.01.017

See Also

[Lemna-models](#), [Macrophyte-models](#), [Transferable](#), [Scenarios](#)

Other Lemna models: [Lemna-models](#), [Lemna_SETAC\(\)](#)

Other macrophyte models: [Lemna_SETAC\(\)](#), [Macrophyte-models](#), [Myrio\(\)](#), [Myrio_log\(\)](#)

Lemna_SETAC

Lemna model (Klein et al. 2021)

Description

The model was described and published by the SETAC Europe Interest Group Effect Modeling (Klein et al. 2022). It is based on the *Lemna* model by Schmitt (2013). The model is a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for the aquatic macrophytes *Lemna spp.*. The model simulates the development of Lemna biomass under laboratory and environmental conditions. Growth of the Lemna population is simulated on basis of photosynthesis and respiration rates which are functions of environmental conditions. The toxicodynamic sub-model describes the effects of growth-inhibiting substances by a respective reduction in the photosynthesis rate based on internal concentrations.

Usage

`Lemna_SETAC()`

Value

an S4 object of type [LemnaSetac](#)

State variables

The model has two state variables:

- BM, Biomass (g dw m⁻²)
- M_int, Mass of toxicant in plant population (mass per m², e.g. ug m⁻²)

Model parameters

- Growth model
 - k_photo_fixed, Model switch for unlimited growth conditions (TRUE/FALSE)
 - k_photo_max, Maximum photosynthesis rate (d-1)
 - k_loss, Reference loss rate (d-1)
 - BM_threshold, Lower biomass abundance threshold, (g dw m-2)
 - BM_min, Reservoir for biomass recovery, (g dw m-2)
- Temperature response of photosynthesis
 - T_opt, Optimum growth temperature (°C)
 - T_min, Minimum growth temperature (°C)
 - T_max, Maximum growth temperature (°C)
- Temperature response of biomass loss rate
 - Q10, Temperature coefficient (-)
 - T_ref, Reference temperature for response=1 (°C)
- Irradiance response of photosynthesis
 - alpha, Slope of irradiance response (m² d kJ⁻¹)
 - beta, Intercept of irradiance response (-)
- Nutrient response of photosynthesis
 - N_50, Half-saturation constant of Nitrogen (mg N L⁻¹)
 - P_50, Half-saturation constant of Phosphorus (mg P L⁻¹)
- Density dependence of photosynthesis
 - BM_L, Carrying capacity (g dw m⁻²)
- Concentration response (Toxicodynamics)
 - EC50_int, Internal concentration resulting in 50% effect (ug L⁻¹)
 - E_max, Maximum inhibition (-)
 - b, Slope parameter (-)
- Internal concentration (Toxicokinetics)
 - P, Permeability (cm d⁻¹)
 - r_A_DW, Area per dry-weight ratio (cm² g⁻¹)
 - r_FW_DW, Fresh weight per dry weight ratio (-)
 - r_FW_V, Fresh weight density (g cm⁻³)
 - r_DW_FN, Dry weight per frond ratio (g dw)
 - K_pw, Partitioning coefficient plant:water (-)
 - k_met, Metabolisation rate (d⁻¹)

Forcings

Besides exposure, the model requires four environmental properties as time-series input:

- tmp, temperature (°C)
- irr, irradiance (kJ m⁻² d⁻¹)

- P, Phosphorus concentration (mg P L⁻¹)
- N, Nitrogen concentration (mg N L⁻¹)

Forcings time-series are represented by `data.frame` objects consisting of two columns. The first for time and the second for the environmental factor in question.

Entries of the `data.frame` need to be ordered chronologically. A time-series can consist of only a single row; in this case it will represent constant environmental conditions. See [scenarios](#) for more details.

Effects

Supported effect endpoints include *BM* (biomass) and *r* (average growth rate during simulation). The effect on biomass is calculated from the last state of a simulation. Be aware that endpoint *r* is incompatible with biomass transfers.

Simulation output

For reasons of convenience, the return value contains by default two additional variables derived from simulation results: the internal concentration `C_int` as well as the number of fronds `FronNo`. These can be disabled by setting the argument `nout = 0`.

The available output levels are as follows:

- `nout >= 1`: `C_int`, internal concentration (mass per volume)
- `nout >= 2`: `FronNo`, frond number (-)
- Response functions
 - `nout >= 3`: `f_loss`, respiration dependency function (-)
 - `nout >= 4`: `f_photo`, photosynthesis dependency function (-)
 - `nout >= 5`: `fT_photo`, temperature response of photosynthesis (-)
 - `nout >= 6`: `fI_photo`, irradiance response of photosynthesis (-)
 - `nout >= 7`: `fP_photo`, phosphorus response of photosynthesis (-)
 - `nout >= 8`: `fN_photo`, nitrogen response of photosynthesis (-)
 - `nout >= 9`: `fBM_photo`, density response of photosynthesis (-)
 - `nout >= 10`: `fCint_photo`, concentration response of photosynthesis (-)
- Environmental variables
 - `nout >= 11`: `C_int_unb`, unbound internal concentration (mass per volume)
 - `nout >= 12`: `C_ext`, external concentration (mass per volume)
 - `nout >= 13`: `Tmp`, temperature (deg C)
 - `nout >= 14`: `Irr`, irradiance (kJ m⁻² d⁻¹)
 - `nout >= 15`: `Phs`, Phosphorus concentration (mg P L⁻¹)
 - `nout >= 16`: `Ntr`, Nitrogen concentration (mg N L⁻¹)
- Derivatives
 - `nout >= 17`: `dBM`, biomass derivative (g dw m⁻² d⁻¹)
 - `nout >= 18`: `dM_int`, mass of toxicant in plants derivative (mass per m² d⁻¹)

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

References

Klein J., Cedergreen N., Heine S., Reichenberger S., Rendal C., Schmitt W., Hommen U., 2021: *Refined description of the Lemna TKTD growth model based on Schmitt et al. (2013) - equation system and default parameters*. Report of the working group *Lemna* of the SETAC Europe Interest Group Effect Modeling. Version 1, uploaded on 22. Sept. 2021. <https://www.setac.org/group/effect-modeling.html>

Schmitt W., Bruns E., Dollinger M., and Sowig P., 2013: *Mechanistic TK/TD-model simulating the effect of growth inhibitors on Lemna populations*. *Ecol Model* 255, pp. 1-10. doi:10.1016/j.ecolmodel.2013.01.017

See Also

[Lemna-models](#), [Macrophyte-models](#), [Transferable](#), [Scenarios](#)

Other Lemna models: [Lemna-models](#), [Lemna_Schmitt\(\)](#)

Other macrophyte models: [Lemna_Schmitt\(\)](#), [Macrophyte-models](#), [Myrio\(\)](#), [Myrio_log\(\)](#)

lik_profile

Likelihood profiling

Description

[Experimental]

The aim of the function is two-fold: 1) estimate a 95% confidence around each parameter of a calibrated model, and 2) see if perhaps a local minimum was found rather than a global minimum. To achieve this, the likelihood profiling goes through every parameter one by one. For each parameter, the model is sequentially refit with the parameter value set to increasingly lower and higher values, and the likelihood of the model given the data calculated (using `log_lik()`). The likelihood is then compared to the likelihood of the original model (using a likelihood ratio). This leads to the development of a likelihood profile, from which a plot a 95% confidence interval for the parameter is derived.

The idea of the function is a variable stepwise algorithm: When the likelihood ratio changes very little (less than `l_crit_min`), the stepsize is increased (up to a maximum, specified by `f_step_max`). When the lik. ratio changes too much (more than `l_crit_max`), the algorithm tries again with a smaller stepsize (also bound to a minimum: `f_step_min`). Note that the stepsize is used as a fraction of the parameter value that is tried. To prevent very small stepsizes when the value goes towards zero (as can be the case for effect thresholds), an absolute minimum stepsize (`f_step_abs`), which is specified as a fraction of the best parameter value (\hat{x}) (unless it is zero, then algorithm takes something small).

The function was inspired by a MatLab BYOM v.6.8 procedure, created by Tjalling Jager. For details, please refer to BYOM (<http://debtox.info/byom.html>) as well as Jager (2021).

Usage

```
lik_profile(
  x,
  par,
  output,
  data = NULL,
  bounds = NULL,
  refit = TRUE,
  type = c("coarse", "fine"),
  break_prof = FALSE,
  ...
)
```

Arguments

<code>x</code>	either a single scenario or a list of caliset objects
<code>par</code>	named vector - parameters (names and values) to be profiled
<code>output</code>	character vector, name of output column of simulate() that is used in calibration
<code>data</code>	only needed if <code>x</code> is a scenario
<code>bounds</code>	optional list of lists (including lower and upper bound): uses defaults in <code>x</code> object, but can be overwritten here (e.g. <code>bounds <- list(k_resp = list(0,10), k_phot_max = list(0,30))</code>)
<code>refit</code>	if 'TRUE' (default), refit if a better minimum is found
<code>type</code>	"fine" or "coarse" (default) likelihood profiling
<code>break_prof</code>	if 'TRUE' (default), stop the profiling if a better optimum is located
<code>...</code>	additional parameters passed on to stats::optim() and calibrate()

Value

A list containing, for each parameter profiled, the likelihood profiling results as a dataframe; the 95% confidence interval; the original parameter value; the likelihood plot object; and the recalibrated parameter values (in case a lower optimum was found)

References

Jager T, 2021. Robust Likelihood-Based Optimization and Uncertainty Analysis of Toxicokinetic-Toxicodynamic Models. *Integrated Environmental Assessment and Management* 17:388-397. doi:10.1002/ieam.4333

Examples

```
# Example with Lemna model - physiological params
library(dplyr)

# exposure - control run
exp <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(time=t, conc)

# observations - control run
obs <- Schmitt2013 %>%
  filter(ID == "T0") %>%
  select(t, BM=obs)

# update metsulfuron
myscenario <- metsulfuron %>%
  set_param(c(k_phot_fix = TRUE, Emax = 1)) %>%
  set_init(c(BM = 12)) %>%
  set_exposure(exp)

fit <- calibrate(
  x = myscenario,
  par = c(k_phot_max = 1),
  data = obs,
  output = "BM",
  lower=0,
  upper=1,
  method="Brent"
)

# Likelihood profiling

res <- lik_profile(
  x = myscenario,
  data = obs,
  output = "BM",
  par = fit$par,
  bounds = list(
    k_phot_max = list(0, 30)
  ),
  refit = FALSE,
  type = "fine",
  method = "Brent"
)

# plot
plot_lik_profile(res)
```

log_enable	<i>Start and stop logging</i>
------------	-------------------------------

Description

Start and stop logging

Usage

```
log_enable(file = NULL, append = TRUE, envir = parent.frame())
```

```
log_disable()
```

Arguments

file	character, file name or path to a log file
append	logical, if TRUE output will be appended to an existing log file, otherwise the log file will be replaced
envir	log will be automatically disabled if environment is exited, set to NULL to disable

Value

no return value

log_envir	<i>Log R environment properties</i>
-----------	-------------------------------------

Description

Log R environment properties

Usage

```
log_envir()
```

Value

no return value

log_lik	<i>Calculate log likelihood</i>
---------	---------------------------------

Description

Calculates the sum of log likelihoods of each observation given the model parameterization (considering a normal distribution around the prediction for each datapoint)

Usage

```
log_lik(npars, obs, pred)
```

Arguments

npars	named numeric vector of parameters that the model was calibrated on
obs	numeric vector of observed values
pred	numeric vector of predicted values

Value

the log likelihood value

Examples

```
# observations
obs <- c(12, 38, 92, 176, 176, 627, 1283, 2640)
# intercept, a, and slope, b, of a Poisson regression fitted through obs
pars <- c(a = 2, b = 0.73)
# predictions with the Poisson regression
pred <- c(15.43, 32.15, 66.99, 139.57, 290.82, 605.94, 1262.52, 2630.58)
# example plot
plot(seq(1:length(obs)), obs)
lines(seq(1:length(obs)), pred)
log_lik(
  npars = length(pars),
  obs = obs,
  pred = pred
)
```

log_msg	<i>Add a log message</i>
---------	--------------------------

Description

Message will only appear in the console or in log file if logging was enabled using `log_enable()`.

Usage

```
log_msg(...)
```

Arguments

... elements will be concatenated using `paste0()`

Value

no return value

Examples

```
log_msg("this message will not appear")

log_enable()
log_msg("this message will appear")
log_msg("a number of ", "elements to ", 42, " concatenate")
```

log_scenarios	<i>Log scenario properties</i>
---------------	--------------------------------

Description

Log scenario properties

Usage

```
log_scenarios(x, header = TRUE)
```

Arguments

x vector of `EffectScenario` objects
header logical, if TRUE a header line will be printed

Value

unmodified argument x

Macrophyte-models	<i>Macrophyte models</i>
-------------------	--------------------------

Description

Population models of standard test macrophytes, such as *Lemna spp.*

Details

Available macrophyte models:

- [Lemna](#)
- [Myriophyllum](#)

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

See Also

[Scenarios](#)

Other macrophyte models: [Lemna_SETAC\(\)](#), [Lemna_Schmitt\(\)](#), [Myrio\(\)](#), [Myrio_log\(\)](#)

Other models: [Algae-models](#), [DEB-models](#), [GUTS-RED-models](#), [Lemna-models](#), [Myriophyllum-models](#)

metsulfuron	<i>Lemna data published by Schmitt (2013)</i>
-------------	---

Description

Data set for the parametrisation of a mechanistic combined toxicokinetic-toxicodynamic (TK/TD) and growth model for the aquatic macrophytes *Lemna spp.* as published by Schmitt *et al.* (2013). The growth model was parameterised by Schmitt *et al.* based on these data while toxicokinetic and toxicodynamic parameters were determined by calibrating the model using substance specific effect data of metsulfuron-methyl.

Usage

metsulfuron

Format

An object of class LemnaSchmittScenario of length 1.

References

Schmitt W., Bruns E., Dollinger M., and Sowig P., 2013: *Mechanistic TK/TD-model simulating the effect of growth inhibitors on Lemna populations*. Ecol Model 255, pp. 1-10. doi:10.1016/j.ecolmodel.2013.01.017

See Also

[Lemna-models](#)

minnow_it

A GUTS-RED-IT scenario of the fathead minnow

Description

The scenario consists of a parameterized GUTS-RED-IT model and a constant exposure series. Model parameters were derived from an acute fish toxicity study of the fathead minnow and chlorpyrifos by Geiger *et al.* (1988). The dataset is also referred to as *GUTS Ring-test dataset C* by EFSA (2018). Fitted parameters were estimated using the *morse* package.

Usage

minnow_it

Format

An object of class GutsRedIt of length 1.

Details

The background mortality rate (hb) was set to zero.

Source

<https://mosaic.univ-lyon1.fr/guts>

References

Geiger D.L., Call D.J., and Brooke L.T., 1988: *Acute toxicities of organic chemicals to fathead minnows (Pimephales promelas): Volume IV*, pp. 195-197. University of Wisconsin-Superior, Center for Lake Superior Environmental Studies. ISBN 9780961496838.

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

[GUTS-RED-models](#)

minnow_sd

A GUTS-RED-SD scenario of the fathead minnow

Description

The scenario consists of a parameterized GUTS-RED-SD model and a constant exposure series. Model parameters were derived from an acute fish toxicity study of the fathead minnow and chlorpyrifos by Geiger *et al.* (1988). The dataset is also referred to as *GUTS Ring-test dataset C* by EFSA (2018). Fitted parameters were estimated using the *morse* package.

Usage

minnow_sd

Format

An object of class GutsRedSd of length 1.

Details

The background mortality rate (hb) was set to zero.

Source

<https://mosaic.univ-lyon1.fr/guts>

References

Geiger D.L., Call D.J., and Brooke L.T., 1988: *Acute toxicities of organic chemicals to fathead minnows (Pimephales promelas): Volume IV*, pp. 195-197. University of Wisconsin-Superior, Center for Lake Superior Environmental Studies. ISBN 9780961496838.

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

[GUTS-RED-models](#)

Myrio

Myriophyllum model with exponential growth

Description

The *Myriophyllum* model is derived from the *Lemna* TKTD model by Klein *et al.* (2021). The *Myriophyllum* model is mathematically equivalent to the Tier 2C version of the *Lemna* model by Klein *et al.* (2021), cf. [Lemna_SETAC\(\)](#). Recommended settings for Tier 2C are `k_photo_fixed=TRUE` and `k_resp=0` (Klein *et al.* 2021). In particular, the growth model is a simple exponential growth model, which is considered to be the typical situation for a laboratory macrophyte study. Instead of frond numbers as for *Lemna*, the biomass is also returned as total shoot length (TSL) in simulation results. Consequently, the model has the additional parameter `r_DW_TSL` (dry weight per total shoot length ratio) instead of `r_DW_FN` (dry weight per frond number ratio).

Usage

```
Myrio()
```

Value

an S4 object of type [MyrioExpScenario](#)

State variables

The model has two state variables:

- BM, Biomass (g dw m⁻² for field studies or mg dw for lab)
- M_{int}, Mass of toxicant in plant population (mass per m², e.g. ug m⁻²)

Model parameters

- Growth model
 - `k_photo_max`, Maximum photosynthesis rate (d⁻¹), default: 0.47
- Concentration response (Toxicodynamics)
 - `EC50_int`, Internal concentration resulting in 50% effect (ug L⁻¹)
 - `E_max`, Maximum inhibition (-), default: 1
 - `b`, Slope parameter (-)
- Internal concentration (Toxicokinetics)
 - `P`, Permeability (cm d⁻¹)
 - `r_A_DW`, Area per dry-weight ratio (cm² g⁻¹), default: 1000
 - `r_FW_DW`, Fresh weight per dry weight ratio (-), default: 16.7
 - `r_FW_V`, Fresh weight density (g cm⁻³), default: 1

- r_{DW_TSL} , Dry weight per total shoot length ratio (g (field) or mg (lab) dw cm⁻¹)
- K_{pw} , Partitioning coefficient plant:water (-), default: 1
- k_{met} , Metabolisation rate (d⁻¹), default: 0

Environmental factors

None.

Parameter boundaries

Default values for parameter boundaries are set for all parameters by expert judgement, for calibration purposes. Values can be modified using `set_bounds()`.

Simulation output

Simulation results will contain the state variables. It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`. As an example, set `nout=2` to enable reporting of the acceleration factor (MV) and the mobilization flux (pC). Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- `nout >= 1`: C_{int} , internal concentration (mass per volume)
- `nout >= 2`: TSL, total shoot length (?)
- `nout >= 3`: f_{photo} , photosynthesis dependency function (-)
- Growth and TK/TD
 - `nout >= 4`: C_{int_unb} , unbound internal concentration (mass per volume)
 - `nout >= 5`: C_{ext} , external concentration (mass per volume)
- Derivatives
 - `nout >= 6`: dB_M , biomass derivative (g dw m⁻² d⁻¹)
 - `nout >= 7`: dM_{int} , mass of toxicant in plants derivative (mass per m² d⁻¹)

Effects

Supported effect endpoints include BM (biomass) and r (average growth rate during simulation). The effect on biomass is calculated from the last state of a simulation. Be aware that endpoint r is incompatible with biomass transfers.

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g.

reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

References

Klein J., Cedergreen N., Heine S., Reichenberger S., Rendal C., Schmitt W., Hommen U., 2021: *Refined description of the Lemna TKTD growth model based on Schmitt et al. (2013) - equation system and default parameters*. Report of the working group *Lemna* of the SETAC Europe Interest Group Effect Modeling. Version 1, uploaded on 22. Sept. 2021. <https://www.setac.org/group/effect-modeling.html>

See Also

[Macrophyte-models](#), [Transferable](#), [Scenarios](#)

Other Myriophyllum models: [Myrio_log\(\)](#), [Myriophyllum-models](#)

Other macrophyte models: [Lemna_SETAC\(\)](#), [Lemna_Schmitt\(\)](#), [Macrophyte-models](#), [Myrio_log\(\)](#)

Myriophyllum-models *Myriophyllum models*

Description

Supported models:

- [Myrio\(\)](#), with exponential growth
- [Myrio_log\(\)](#), with logistic growth

See Also

[Lemna-models](#), [Transferable](#)

Other Myriophyllum models: [Myrio\(\)](#), [Myrio_log\(\)](#)

Other models: [Algae-models](#), [DEB-models](#), [GUTS-RED-models](#), [Lemna-models](#), [Macrophyte-models](#)

Myrio_log

Myriophyllum model with logistic growth**Description**

The *Myriophyllum* model is derived from the *Lemna* TKTD model by Klein *et al.* (2021). `Myrio_log()` modifies the `Myrio()` model to feature logistic growth, i.e. control growth is described by the differential equation $d\text{ BM}/dt = k_{\text{photo_max}} \cdot \text{BM} \cdot (1 - \text{BM}/\text{BM_L})$ where `BM_L` is the carrying capacity.

Usage

```
Myrio_log()
```

Value

an S4 object of type `MyrioLogScenario`

Model parameters

- Growth model
 - `k_photo_max`, Maximum photosynthesis rate (d-1), default: 0.47
 - `BM_L`, Carrying capacity (g dw m-2)
- Concentration response (Toxicodynamics)
 - `EC50_int`, Internal concentration resulting in 50% effect (ug L-1)
 - `E_max`, Maximum inhibition (-), default: 1
 - `b`, Slope parameter (-)
- Internal concentration (Toxicokinetics)
 - `P`, Permeability (cm d-1)
 - `r_A_DW`, Area per dry-weight ratio (cm² g-1), default: 1000
 - `r_FW_DW`, Fresh weight per dry weight ratio (-), default: 16.7
 - `r_FW_V`, Fresh weight density (g cm-3), default: 1
 - `r_DW_TSL`, Dry weight per total shoot length ratio (?)
 - `K_pw`, Partitioning coefficient plant:water (-), default: 1
 - `k_met`, Metabolisation rate (d-1), default: 0

State variables

The model has two state variables:

- `BM`, Biomass (g dw m-2 for field studies or mg dw for lab)
- `M_int`, Mass of toxicant in plant population (mass per m², e.g. ug m-2)

Environmental factors

None.

Simulation output

Simulation results will contain the state variables. It is possible to amend the output of `simulate()` with additional model quantities that are not state variables, for e.g. debugging purposes or to analyze model behavior. To enable or disable additional outputs, use the optional argument `nout` of `simulate()`. As an example, set `nout=2` to enable reporting of the acceleration factor (MV) and the mobilization flux (pC). Set `nout=0` to disable additional outputs (default).

The available output levels are as follows:

- `nout >= 1`: `C_int`, internal concentration (mass per volume)
- `nout >= 2`: TSL, total shoot length (?)
- `nout >= 3`: `f_photo`, photosynthesis dependency function (-)
- Growth and TK/TD
 - `nout >= 4`: `C_int_unb`, unbound internal concentration (mass per volume)
 - `nout >= 5`: `C_ext`, external concentration (mass per volume)
- Derivatives
 - `nout >= 6`: `dBm`, biomass derivative (g dw m⁻² d⁻¹)
 - `nout >= 7`: `dM_int`, mass of toxicant in plants derivative (mass per m² d⁻¹)

Effects

Supported effect endpoints include *BM* (biomass) and *r* (average growth rate during simulation). The effect on biomass is calculated from the last state of a simulation. Be aware that endpoint *r* is incompatible with biomass transfers.

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using `set_transfer()`.

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

Parameter boundaries

Default values for parameter boundaries are set for all parameters by expert judgement, for calibration purposes. Values can be modified using `set_bounds()`.

References

Klein J., Cedergreen N., Heine S., Reichenberger S., Rendal C., Schmitt W., Hommen U., 2021: *Refined description of the Lemna TKTD growth model based on Schmitt et al. (2013) - equation system and default parameters*. Report of the working group *Lemna* of the SETAC Europe Interest Group Effect Modeling. Version 1, uploaded on 22. Sept. 2021. <https://www.setac.org/group/effect-modeling.html>

See Also

[Transferable, Scenarios](#)

Other Myriophyllum models: [Myrio\(\)](#), [Myriophyllum-models](#)

Other macrophyte models: [Lemna_SETAC\(\)](#), [Lemna_Schmitt\(\)](#), [Macrophyte-models](#), [Myrio\(\)](#)

no_exposure

Zero exposure

Description

Creates an [ExposureSeries](#) with zero concentration. When setting the zero exposure, pay attention not to accidentally reset the output times of your scenario as the zero exposure series contains only a single time point. See the examples.

Usage

```
no_exposure()
```

Value

an S4 object of type [ExposureSeries](#)

See Also

[set_noexposure\(\)](#)

Examples

```
# this will reset the output times of the sample scenario,
# simulate() will quit with an error
try(
  minnow_it %>%
    set_exposure(no_exposure()) %>%
    simulate()
)

# set zero exposure, but keep original output times
minnow_it %>%
  set_exposure(no_exposure(), reset_times=FALSE) %>%
  simulate()
```

parameter_set	<i>Set of model parameters</i>
---------------	--------------------------------

Description

Set of model parameters

Usage

```
parameter_set(model, param = list(), tag = NA_character_)
```

Arguments

model	character, a string containing a model name, e.g. "GUTS-RED-IT"
param	named list of model parameters
tag	character, an optional identifier

Value

an S4 object of type [ParameterSet](#)

Slots

model	character, a string containing a model name, e.g. "GUTS-RED-IT"
tag	character, an optional identifier
param	named list of model parameters

Examples

```
# create a parameter set and assign it
ps <- parameter_set("GUTS-RED-IT", list(kd=0.12, hb=0.3))
GUTS_RED_IT() %>% set_param(ps)

# multiple scenarios can be modified at once
c(GUTS_RED_IT(), GUTS_RED_IT()) %>%
  set_param(ps)

# model names must match, otherwise an error will be raised
try(GUTS_RED_SD() %>% set_param(ps))
```

pll_debug	<i>Disable parallelization for debugging</i>
-----------	--

Description

In certain cases it might be beneficial to disable parallel execution of e.g. effect profile calculations. By disabling, all processes run sequentially and instantly pass messages to the console which would be delayed during parallel processing. This makes it easier to pinpoint problems within the data or algorithm.

Usage

```
pll_debug(state = TRUE)
```

Arguments

state	logical, if TRUE then parallelization is disabled
-------	---

Value

no return value

plot	<i>S3 plotting functions</i>
------	------------------------------

Description

These functions overload `base::plot()` to provide simple plotting routines to display various time-series and scenario objects.

Usage

```
## S3 method for class 'cvasi.drc'
plot(x, y, scale_x = c("auto", "log10", "none"), ...)
```

```
## S3 method for class 'cvasi.simulate'
plot(x, y, ...)
```

Arguments

x	object to plot
y	unused parameter
scale_x	character, controls how the x-axis is scaled. log10 for a log10-scaled axis, none for no scaling, and auto for automatic selection
...	unused parameters

Value

ggplot2 plot object

Methods (by class)

- `plot(cvasi.drc)`: Plot dose response curves
- `plot(cvasi.simulate)`: Plot return value of `simulate()`

plot_epx

Plot EPx values

Description

[Experimental]

Usage

```
plot_epx(
  EPx_ts,
  exposure_ts,
  draw = TRUE,
  time_col = "time",
  conc_col = "conc",
  epx_x_title = "Start time",
  conc_y_title = "Exposure conc."
)
```

Arguments

<code>EPx_ts</code>	the result of <code>epx_mtw</code> , ie. a tibble with <code>window.start</code> , <code>window.end</code> , <code>endpoint</code> , <code>level</code> and <code>EPx</code>
<code>exposure_ts</code>	an exposure time series with columns for time 't' and concentration 'conc'
<code>draw</code>	Should the whole plot be drawn? If <code>FALSE</code> the exposure plot and the EPx plot are returned as a list for later modification
<code>time_col</code>	the name of the time column in the exposure dataset
<code>conc_col</code>	the name of the concentration column in the exposure dataset
<code>epx_x_title</code>	title of the x-axis of the epx panel
<code>conc_y_title</code>	title of the y-axis of the concentration panel

Value

a grid of ggplots

Examples

```

ti <- 0:21
expo <- abs(0.01*ti + rnorm(length(ti), 0, 0.05))
exposure <- data.frame(time = ti, conc = expo)
metsulfuron_epx_mtw <- metsulfuron %>%
set_exposure(exposure) %>%
epx_mtw(level = 10, factor_cutoff = 1000)
metsulfuron_epx_mtw
plot_epx(EPx_ts = metsulfuron_epx_mtw,
exposure_ts = exposure, conc_y_title = "env. concentration [ $\mu\text{g/L}$ "])
```

plot_lik_profile *Plot likelihood profiles or all profiled parameters*

Description

The function provides a combined plot of the likelihood profiles of all parameters profiled.

Usage

```
plot_lik_profile(x)
```

Arguments

x object of class lik_profile

Value

plots

plot_param_space *Plot likelihood profiles or all profiled parameters*

Description

The function provides bivariate parameter space plots indicating parameter draws (from the 95% confidence intervals per parameter obtained through likelihood profiling) that fall within the inner rim (in green, i.e. parameter sets which are not significantly different from the original, based on a chi-square test). The original parameter set is also indicated (in orange), and, if different from the original set, the best fit parameter set is indicated (in red)

Usage

```
plot_param_space(x)
```

Arguments

x object of class param_space

Value

plots

plot_ppc *Creates a PPC plot for a single dataset*

Description

[Experimental]

Usage

```
plot_ppc(
  rs_mean,
  rs_range,
  col_number = 2,
  obs_mean = NULL,
  obs_full = NULL,
  xy_lim = NULL,
  study = NULL
)
```

Arguments

rs_mean data.frame, model results best fit params
rs_range data.frame, predictions (min, max from param.sample run)
col_number column to plot, default = 2
obs_mean data.frame, observations with means per treatment level
obs_full data.frame, full data set including results for replicates
xy_lim optional numeric, limits of x and y axis for plotting
study optional string, name of study which can be used as key

Details

A sample of parameters representing the uncertainty within the dataset is passed to the function. All parameter combinations and exposure patterns are simulated and the range of predicted frond numbers is derived for a single study. The uncertainty is displayed by a Posterior Predictive Plot (PPC). The data (rs_mean, obs_mean and obs_full) must have the following format (col1 = time, col2 = data of interest, col3 = trial name). Data for uncertainties (rs_range) must have the format: col1 = time, col2 = lower boundaries, col3 = upper boundaries, col4 = trial. The user should take care of the input data and consider whether control data and data at time zero should be included in the model check.

Value

a ggplot2 plot object

plot_ppc_combi	<i>Create PPC plot for one or more datasets</i>
----------------	---

Description

[Experimental]

Usage

```
plot_ppc_combi(table, xy_lim = NULL)
```

Arguments

table	data.frame containing return values of calls to plot_ppc()
xy_lim	optional numeric, limits of x and y axis for plotting

Details

The function expects a data.frame with five mandatory and one optional column. The mandatory columns are as follows:

- pred: mean of predictions e.g. frond number for lemna
- max: maximum of predictions
- min: minimum of predictions
- obs: observations
- PPC: color code The optional column is to be named study and contains a study identifier. If more than one study identifier is present in the table, individual studies will be plotted in different colors and a legend will be displayed. The function is called by plot_ppc where the column names are defined (see rs_ppc object).

Value

a ggplot2 plot object

plot_scenario	<i>Creates a prediction plot for one effect scenario</i>
---------------	--

Description

[Deprecated]

Usage

```
plot_scenario(model_base, plot_col = 2, trial_number = NULL)
```

Arguments

model_base	effect scenario object with mean parameters
plot_col	output column which should be plotted, default = 2
trial_number	name for model run (if available tag is used)

Details

This function has been deprecated and replaced by the generic `plot()`.

Sometimes it is helpful if the user can plot results of one effect scenario. This is for instance the case for test simulations or predictions for one profile. This function runs the simulation for one effect scenario and plots the results. Function plots the time (column 1) and the predictions (column 2, can be changed by the user `plot_col`)

Value

plot of the results for one effect scenario

Examples

```
# Please use `plot()` instead
metsulfuron %>%
  simulate() %>%
  plot()
```

plot_sd	<i>Creates plot of model results (uncertainties optional)</i>
---------	---

Description

[Experimental]

Usage

```
plot_sd(
  model_base,
  treatments,
  rs_mean,
  rs_range = NULL,
  obs_mean = NULL,
  obs_full = NULL,
  x_breaks = NULL,
  y_lim = NULL,
  grid_labels = NULL,
  grid_ncol = 2,
  plot_col = 2,
  y_title = NULL,
  ...
)
```

Arguments

model_base	effect scenario object with mean parameters
treatments	treatments exposure levels as data frame
rs_mean	data.frame, model results best fit params
rs_range	data.frame, uncertainties as data frame
obs_mean	data.frame, observation data with means per treatment level
obs_full	data.frame, full set including results for replicates
x_breaks	optional vector of breaks of x-axis
y_lim	optional vector containing limits of y-axis
grid_labels	optional labels of grid headers
grid_ncol	optional number of grid columns
plot_col	output column which should be plotted
y_title	optional title of y-axis
...	any additional parameters

Details

All parameter combinations and exposure patterns are simulated and the mean of predictions is derived for a single study. The uncertainty is passed to the function due to computation time. Results are displayed by plotting the time series including the uncertainty interval. Observation data can be optionally displayed. Data should be provided in long format. Function plots the time (column 1) and the predictions (column 2, can be changed by the user plot_col)

Value

a ggplot2 plot object

Examples

```
set.seed(124)
exposure <- data.frame(
  time = 0:21,
  conc = rnorm(n = 22, mean = 0.1, sd = 0.06),
  trial = "T1"
)
forcings <- list(temp = 12, rad = 15000)
param <- list(EC50 = 0.3, b = 4.16, P_up = 0.0054)
inits <- list(BM = 0.0012, E = 1, M_int = 0)

scenario <- Lemna_Schmitt() %>%
  set_forcings(forcings) %>%
  set_param(param) %>%
  set_init(inits)

sim_result <- simulate_batch(
  model_base = scenario,
  treatments = exposure,
  param_sample = NULL
)

plot_sd(
  model_base = scenario,
  treatments = exposure,
  rs_mean = sim_result
)
```

pull_metadata

Pull metadata from scenarios

Description

The method pulls available metadata from scenario objects and returns a table with additional columns. If the argument already was a data.frame object, the columns are appended. May overwrite existing columns of the same name.

Usage

```
pull_metadata(x, model = TRUE, exposure = TRUE)
```

Arguments

x	vector of scenarios or a data.frame containing a column scenario with EffectScenario objects
model	logical, if TRUE then model metadata is pulled
exposure	logical, if TRUE then exposure series metadata is pulled

Value

a data.frame

Examples

```
metsulfuron %>%  
  pull_metadata()
```

Rsubcapitata	<i>An algae scenario</i>
--------------	--------------------------

Description

Data are from Weber 2012 publication.

Usage

```
Rsubcapitata
```

Format

An object of class `AlgaeTKTD` of length 1.

References

Weber D, Schaeffer D, Dorgerloh M, Bruns E, Goerlitz G, Hammel K, Preuss TG and Ratte HT, 2012. Combination of a higher-tier flow-through system and population modeling to assess the effects of time-variable exposure of isotroturon on the green algae *Desmodesmus subspicatus* and *Pseudokirchneriella subcapitata*. *Environmental Toxicology and Chemistry*, 31, 899-908. [doi:10.1002/etc.1765](https://doi.org/10.1002/etc.1765)

See Also

[Algae_TKTD](#)

Description

The `EffectScenario` class is the base for all of the basic scenario types and models. It contains slots for data and settings that are required by most models such as a vector of model parameters and a vector of initial states. For each particular model, the class's slots are filled with certain default or fixed values. Some models derive from this class and add slots to store additional data.

Details

Certain behaviors that are required to model complex processes cannot be represented by a single `EffectScenario`. As an example, the parameters of a scenario are generally fixed during the simulated time period. In order to represent a change in parameter values, the original scenario would need to split into two scenarios *A* and *B* which differ by parameter values and simulated time period. By combining these scenarios to a *scenario sequence*, the sequence would be treated as a single, complex scenario. See [sequence\(\)](#) for more information.

Parameters:

Most parameters are represented by numerical types but other types are possible depending on model. Please refer to the model description which parameters are required and in which unit. Some or all parameters may be required to start a simulation. If required parameters are missing, simulation will fail with an error message.

Initial state:

The *initial state* represents the starting values of state variables when starting a simulation. A scenario's default initial state may be insufficient to get sensible results. It is advisable to set an initial state explicitly when creating a new scenario, see [set_init\(\)](#).

In theory, a scenario's state variables can be renamed by modifying the names of the initial state vector. However, this is strongly discouraged as this will affect other routines such as [effect\(\)](#) and [epx\(\)](#) and may render results useless.

Exposure:

Exposure refers to the concentration of toxicant an organism is exposed to. In case of aquatic organisms, this would commonly be the concentration of a toxicant in water. Other interpretations are possible depending on model assumptions.

Exposure time-series are generally represented by a `data.frame` containing two columns. The first column representing time, the second representing the exposure level. The ordering of columns is mandatory. The column names are essentially irrelevant but sensible names may help documenting the scenario and its data. The rows must be ordered chronologically. A time-series can consist of only a single row; in this case it will represent constant exposure. Exposure time-series are set to a scenario using [set_exposure\(\)](#).

Handling time-series is a costly task for the ODE solver due to consistency checks and interpolation between time steps. How the solver interpolates the time-series can be controlled by certain arguments to functions such as [simulate\(\)](#) and [effect\(\)](#). Please refer to [simulate\(\)](#) for a brief overview and [deSolve::forcings](#) for a detailed description.

Exposure time-series should be kept as short as possible and as complex as needed for optimal computational efficiency.

Environmental forcings:

Forcings generally refer to model parameters that change over time as part of an external function such as environmental temperature and exposure levels. Due to the importance of exposure in regulatory assessments, this R package explicitly distinguishes between environmental forcings and exposure. However, the same restrictions and features apply to both of them.

Forcing time-series are handled the same way as exposure time-series, i.e. they are represented by a `data.frame` containing two columns. The first column representing time, the second representing the parameter that is a function of time. The ordering of columns is mandatory. The rows must be ordered chronologically. Forcings time-series are set using `set_forcings()`. Please refer to the *Exposure* section for more information on how time-series are handled.

Output times:

A scenario's simulated time period is defined by its minimum and maximum output time. Simulation results will only be returned for the defined output times even though the ODE solver may use smaller time steps between output times. Output times can be explicitly set using `set_times()`. The number and distance of output times may have influence on the precision of simulation results and numerical stability, cf. `simulate()`.

Be aware that `set_exposure()` will overwrite previously defined output times if not requested otherwise.

Effects:

Generally, all state variables can be used as effect endpoints but models may provide additional endpoints. Use `set_endpoints()` to enable or disable endpoints for a scenario.

Some scenarios or models require control runs to calculate effects under exposure. Generally, control simulations will run automatically where needed. However, when conducting a large number of repeated simulations, e.g. when calculating effect profiles (EPx values) or simulating moving exposure windows, it may be computationally efficient to run control simulations only once and cache their results within the scenario. Please refer to `cache_controls()` for details.

Moving exposure windows:

The time frame relevant for effects may be much shorter than the assessed exposure time-series for certain organisms. This fact can be represented by moving exposure windows which divide a long time period in a number of consecutive windows of the same length. Each window is simulated individually and effects are calculated. By default, methods such as `effect()` will only return the maximum effect of all considered windows but detailed results can be presented on demand.

To use moving exposure windows, the exposure time-series must be regular, i.e. must have an equidistant step length in time. The length of the window is defined as the number of time steps of the exposure time-series. As an example, assume the time-series has daily granularity and a moving window of seven days length is required. In this case, the moving window must have a length of seven (7) time steps. If the exposure time-series had hourly granularity, the same window would need to have a length of 168 (=7*24) time steps. Please refer to `set_window()` for details.

Slots

name character, unique model name
 tag character, an optional identifier
 param list of parameter key-value pairs
 param.bounds named list of parameter boundaries
 param.req character vector of required parameters
 forcings list of data.frames representing forcing time-series
 forcings.req character vector or required model forcings data, e.g. temperature
 init list of initial model states
 times numeric vector of output times, beginning and end also define the simulated period
 endpoints character vector of endpoints to calculate results for
 exposure data.frame with two columns representing an exposure time-series
 control list of named numerical vectors, contains the control values for all relevant moving windows
 control.req logical, if TRUE then control values are required to calculate effects
 window.length numeric, maximum length of the simulated period, if window.length is shorter than the exposure pattern, then all possible exposure sub-patterns are evaluated for effect calculation. This is also referred to as a moving window approach.
 window.interval numeric, interval determining distance between moving windows during effect calculation. First window starts at first time point in exposure pattern.

See Also

Other scenario-related: [Transferable](#)

 Schmitt2013

A Lemna data set with multiple treatment levels

Description

Data are from Schmitt 2013 publication.

Usage

```
Schmitt2013
```

Format

An object of class data.frame with 56 rows and 4 columns.

References

Schmitt W., Bruns E., Dollinger M., and Sowig P., 2013: *Mechanistic TK/TD-model simulating the effect of growth inhibitors on Lemna populations*. Ecol Model 255, pp. 1-10. doi:10.1016/j.ecolmodel.2013.01.017

See Also

[Lemna-models](#)

sequence	<i>Sequence of scenarios</i>
----------	------------------------------

Description

A sequence of scenarios is treated as a single scenario and each scenario is simulated one after the other. If scenario n in a sequence was simulated, scenario $n+1$ will start off in the model state where n had ended. Scenario sequences can be used to e.g. implement changes in model parameters over time.

Details

Requirements:

All scenarios in a sequence must fulfill the following requirements:

- All scenarios must have identical state variables
- The *output times* of all scenarios must represent a continuous time series without gaps or overlaps

Limitations:

Only simulation of sequences are supported, at the moment. Effects and effect profiles (EPx values) cannot be derived, yet.

Value

an S4 object of type [ScenarioSequence](#)

Examples

```
# create two scenarios that need to be simulated one after the other
scen1 <- minnow_it %>% set_times(0:3)
scen2 <- minnow_it %>% set_times(3:6) %>% set_param(c(kd=0))

# create a sequence and assign scenarios
sq <- sequence(list(scen1, scen2))

# simulate the sequence

simulate(sq)
```

set_bounds	<i>Set boundaries of model parameters</i>
------------	---

Description

Modifies the boundaries of model parameters for one or more [scenario](#) or [caliset](#) objects.

Usage

```
set_bounds(x, bounds)

## S4 method for signature 'EffectScenario,list'
set_bounds(x, bounds)

## S4 method for signature 'CalibrationSet,list'
set_bounds(x, bounds)

## S4 method for signature 'list,list'
set_bounds(x, bounds)
```

Arguments

x	vector of scenario or caliset objects
bounds	named list of numerical vectors, where the first level lists the parameters by name, and the second level lists the lower and upper boundary

Value

[scenario](#) or [caliset](#) with modified parameter boundaries

Examples

```
metsulfuron %>%
  set_bounds(list(k_phot_max = c(0, 30),
                 k_resp = c(0, 10)))
```

set_endpoints	<i>Set effect endpoints</i>
---------------	-----------------------------

Description

Effect endpoints calculated by functions such as [effect\(\)](#) and [epx\(\)](#) can be enabled and disabled. If an endpoint is not required for an assessment, it should be disabled for reasons of computational efficiency. Please refer to the model description for a list of available endpoints.

Usage

```
set_endpoints(x, endpoints)
```

Arguments

```
x                vector of EffectScenario objects
endpoints        character vector of endpoint names
```

Value

Modified EffectScenario objects

Examples

```
# Only enable reproduction (R) endpoint for americamysis scenario
americamysis %>%
  set_endpoints("R") %>%
  effect()

# Enable endpoints length (L) and reproduction (R)
americamysis %>%
  set_endpoints(c("L", "R")) %>%
  effect()
```

set_exposure	<i>Set exposure time-series</i>
--------------	---------------------------------

Description

Exposure refers to the toxicant concentration an organism is exposed to. In case of aquatic organisms, this would commonly be the concentration of a toxicant in water. Other interpretations are possible depending on model assumptions.

Usage

```
set_exposure(scenarios, series, ...)
```

```
## S4 method for signature 'ANY,ANY'
```

```
set_exposure(scenarios, series)
```

```
## S4 method for signature 'EffectScenario,data.frame'
```

```
set_exposure(scenarios, series, ...)
```

```
## S4 method for signature 'EffectScenario,ExposureSeries'
```

```
set_exposure(scenarios, series, reset_times = TRUE)
```

```
## S4 method for signature 'EffectScenario,list'
```

```

set_exposure(scenarios, series, ...)

## S4 method for signature 'list,list'
set_exposure(scenarios, series, ...)

## S4 method for signature 'list,ANY'
set_exposure(scenarios, series, ...)

```

Arguments

scenarios	vector of scenarios
series	vector of ExposureSeries objects or a single <code>data.frame</code>
...	additional arguments
reset_times	logical, if TRUE, the exposure time-series' time points will be set as output times. Defaults to TRUE

Details

Exposure time-series are generally represented by a `data.frame` containing two columns. The first column for time, the second representing the exposure level. The ordering of columns is mandatory. The column names are non-relevant but sensible names may help documenting the scenario and its data. The `data.frame`'s rows must be ordered chronologically. A time-series can consist of only a single row; in this case it will represent constant exposure.

For convenience, a time-series with zero exposure can be set using [set_noexposure\(\)](#).

Computational efficiency:

Handling time-series is a costly task for the ODE solver due to consistency checks and interpolation between time steps. How the solver interpolates the time-series can be controlled by optional arguments to functions such as [simulate\(\)](#) and [effect\(\)](#). Please refer to [simulate\(\)](#) for a brief overview and [deSolve::forcings](#) for a detailed description.

Exposure time-series should be kept as short as possible and as complex as needed for optimal computational efficiency.

Output times:

By default, the exposure time-series' time points will also be used as output times of the scenario. Any output times previously set by [set_times\(\)](#) will be lost. If this behavior is undesired, set the function argument `reset_times=FALSE`.

Multiple exposure series and scenarios:

The functions supports modifying multiple scenarios at once: by calling it with lists of [scenario](#) and [ExposureSeries](#) objects. The cartesian product of all scenarios and exposure series will be returned, iff the parameter `expand = TRUE` is set.

As an example for the *expand* mode, two scenarios A and B and one exposure series g will result in two scenarios Ag and Bg, both using exposure series g. Two scenarios A and B as well as two exposure series g and h will result in four scenarios Ag,Ah,Bg, and Bh.

Value

list of `EffectScenario` objects

Examples

```

# set a data.frame as exposure series
Lemna_Schmitt() %>% set_exposure(data.frame(time=c(0, 1, 2, 3), conc=c(1, 1, 0, 0)))

# set one ExposureSeries
es1 <- ExposureSeries(data.frame(time=0, conc=0))
Lemna_Schmitt() %>% set_exposure(es1)

# set two ExposureSeries to create two scenarios
es2 <- ExposureSeries(data.frame(time=5:10, conc=1))
Lemna_Schmitt() %>% set_exposure(c(es1, es2))

# set one ExposureSeries without resetting existing output times
Lemna_Schmitt() %>%
  set_times(0:5) %>%
  set_exposure(es1, reset_times=FALSE)

```

set_forcings

Set time-dependent parameters

Description

Parameters which change their value over time are referred to as *forcings*. If and what parameters can vary over time depends on the model in question. In many cases, *forcings* represent time-series of environmental properties.

Usage

```

set_forcings(x, ...)

## S4 method for signature 'EffectScenario'
set_forcings(x, ...)

## S4 method for signature 'list'
set_forcings(x, ...)

```

Arguments

```

x          (vector of) scenario objects
...       named argument list to set as forcings

```

Details

Forcing time-series are always represented by a `data.frame` containing two columns. The first column representing time, the second representing the parameter that is a function of time. The ordering of columns is mandatory. The column names are essentially irrelevant but may help documenting the scenario and its data. The rows must be ordered chronologically. A time-series can consist of only a single row; in this case it will represent constant conditions.

Handling forcing time-series is a costly task for the ODE solver due to consistency checks and interpolation between timesteps. How the solver interpolates the forcing time-series can be controlled by certain arguments to functions such as `simulate()` and `effect()`. Please refer to `simulate()` for a brief overview and `deSolve::forcings` for a detailed description.

Forcing time-series should be kept as short as possible and as complex as needed for optimal computational efficiency.

Value

Modified `scenarios`

Examples

```
# constant values will be automatically converted to a data.frame
Lemna_Schmitt() %>% set_forcings(temp=20) -> lemna
lemna@forcings

# setting multiple forcings at once
df <- data.frame(t=0:14, temp=rnorm(15, mean=20)) # random temperature series
Lemna_Schmitt() %>% set_forcings(temp=df, rad=15000) -> lemna
lemna@forcings

# forcings can also be supplied as a named list
Lemna_Schmitt() %>% set_forcings(list(temp=20, rad=15000)) -> lemna
lemna@forcings
```

set_init

Set initial state

Description

The *initial state* represents the starting values of a scenario's state variables when starting a simulation. A scenario's default initial state may be insufficient to get sensible results.

Usage

```
set_init(x, init)

## S4 method for signature 'vector'
set_init(x, init)

## S4 method for signature 'EffectScenario'
set_init(x, init)
```

Arguments

`x` vector of `EffectScenario` objects
`init` named numeric vector

Details

In theory, a scenarios's state variables can be renamed by modifying the names of the initial state vector. However, this is strongly discouraged as this will affect other routines such as `effect()` and `epx()` and may render results useless.

Value

modified EffectScenario objects

Examples

```
# Set initial biomass to 1.0
metsulfuron %>% set_init(c(BM=1.0)) %>% simulate()
```

set_mode_of_action	<i>Set mode of action</i>
--------------------	---------------------------

Description

Updates the model parameter MoA to a certain value

Usage

```
set_mode_of_action(x, code)
```

```
set_moa(x, code)
```

Arguments

x vector of [scenarios](#)

code a code for a mode of action, refer to model description for details

Value

modified [scenarios](#)

Functions

- `set_moa()`: Shorthand version

Examples

```
# Set MoA=8, i.e. hazard during oogenesis
americamysis %>%
  set_mode_of_action(8) %>%
  effect(method="ode45")

# alternative approach using the parameter directly
americamysis %>%
  set_param(c(MoA=8)) %>%
  effect(method="ode45")
```

set_noexposure	<i>Set zero exposure</i>
----------------	--------------------------

Description

The scenarios current exposure is replaced by a constant exposure time-series of value zero(0.0). Output times are unaffected.

Usage

```
set_noexposure(x)
```

Arguments

x vector of [scenarios](#)

Value

vector of [scenarios](#)

Examples

```
# Derive effect size in sample scenario without toxicant exposure
minnow_it %>%
  set_noexposure() %>%
  effect()
```

set_param	<i>Set model parameters</i>
-----------	-----------------------------

Description

Modifies the parameters of one or more EffectScenario objects.

Usage

```
set_param(x, param)

## S4 method for signature 'EffectScenario,vector'
set_param(x, param)

## S4 method for signature 'EffectScenario,ParameterSet'
set_param(x, param)

## S4 method for signature 'list,ParameterSet'
set_param(x, param)

## S4 method for signature 'list,vector'
set_param(x, param)

## S4 method for signature 'ScenarioSequence,vector'
set_param(x, param)

## S4 method for signature 'ScenarioSequence,ParameterSet'
set_param(x, param)
```

Arguments

x	object(s) to modify
param	named numeric vector with parameter names and value OR a list of parameter_set objects

Details

Most parameters are represented by numerical types but other types are possible depending on model. Please refer to the model description which parameters are required and in which unit. Some or all parameters may be required to start a simulation. If required parameters are missing, simulation will fail with an error message.

Value

Vector of modified objects

Examples

```
Lemna_Schmitt() %>% set_param(c(Emax=1, EC50=0.12))
```

set_tag

Set a tag

Description

Sets the user-defined, custom tag of a scenario. Tags can be helpful to quickly distinguish scenarios of the same model type.

Usage

```
set_tag(x, tag)
```

Arguments

x (vector of) EffectScenario objects
tag vector of character

Value

(vector of) modified EffectScenario objects

See Also

[get_tag\(\)](#)

Examples

```
# set a custom tag  
myscenario <- GUTS_RED_SD() %>% set_tag("My Custom Tag")  
  
# returns `My Custom Tag`  
get_tag(myscenario)  
  
# the tag also appears in the scenario overview  
myscenario
```

set_times	<i>Set output times</i>
-----------	-------------------------

Description

Minimum and maximum output times define the simulated period for a scenario. Simulation results will be returned for each output time, see [simulate\(\)](#).

Usage

```
set_times(x, times)
```

Arguments

x	vector of scenarios
times	numerical vector

Details

Be aware that output times may be modified by [set_exposure\(\)](#). Precision of simulation results may be influenced by chosen output times, see [simulate\(\)](#) for more information.

Value

Vector of modified [scenarios](#)

See Also

[simulate\(\)](#)

Examples

```
# Set simulated period to [2,4] with output intervals of length 1
minnow_it %>% set_times(c(2,3,4))

# Decrease output interval length to 0.1
minnow_it %>% set_times(seq(2, 4, 0.1))
```

set_transfer	<i>Set transfer events</i>
--------------	----------------------------

Description

A *transfer* refers to an event where a certain amount of biomass (*BM*) is moved to a new medium after a period of time. This feature replicates a procedure occurring e.g. in Lemna effect studies and may be necessary to recreate study results.

Usage

```
set_transfer(x, interval, times, biomass, scaled_comp)
```

```
## S4 method for signature 'ANY'
```

```
set_transfer(x, interval, times, biomass, scaled_comp)
```

```
## S4 method for signature 'Transferable'
```

```
set_transfer(x, interval, times, biomass, scaled_comp)
```

Arguments

x	vector of EffectScenario objects
interval	optional numeric, interval in time units of the scenario, set to -1 to disable transfers.
times	optional numeric vector of time points where transfers occur
biomass	optional numeric vector, amount of biomass that is being transferred at each transfer
scaled_comp	optional character vector of affected compartments that are scaled according to new biomass levels

Details

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

Transferred biomass:

At each transfer, a defined amount of biomass is transferred to a new medium. This is modeled by interrupting the simulation at a transfer time point, modifying the biomass level *BM*, and scaling affected compartments according to new biomass levels. Scaling of compartments depending on biomass, such as *internal toxicant mass*, is necessary to correctly reflect mass balances and concentrations over time.

Transferred biomass is set using the *biomass* parameter. It is either a single numerical value in which case the same biomass level is set at each transfer. Or it is a vector of numerical values with the same length as the *times* parameter in which case a custom biomass level can be set for each transfer. Multiple biomass levels can only be set in conjunction with custom transfer time points.

Some scenario types define default values for transferred biomass based on common study set ups.

Regular and custom transfer time points:

Transfers can occur either in regular intervals of time or at selected, custom time points. For regular intervals, the parameter `interval` is set to a single numeric value which has the same unit as the scenario's time dimension. As an example: if a scenario uses the unit of *days* for time, the transfer interval is also specified in *days*:

Transfers occurring at custom time points are set by passing a numerical vector to the parameter `times`. The time points' units must match with the unit of time in the scenario. A custom transfer time point **must not occur at the starting time point of a simulation**.

Affected compartments:

Some compartments depend on biomass to correctly reflect mass balances and concentrations over time, such as *internal toxicant mass*. These compartments need to be scaled linearly to reflect the change in biomass levels. The parameter `scaled_comp` accepts a character vector of compartment names which are scaled at each transfer. This parameter should only be used with custom, user-defined models. If no compartment needs to be scaled, set or use the default value of `character(0)`.

Value

Modified EffectScenario objects

See Also

[Lemna_Schmitt\(\)](#)

Examples

```
# Simulate biomass transfer of 50 *g dw/m2* at a regular interval of 7 *days*
metsulfuron %>%
  set_transfer(interval=7, biomass=50) %>%
  simulate()

# Simulate irregular biomass transfers occurring at days 5, 10, and 12
metsulfuron %>%
  set_transfer(times=c(5,10,12), biomass=50) %>%
  simulate()

# Simulate irregular transfers with changing amounts of transferred biomass
metsulfuron %>%
  set_transfer(times=c(5,10,12), biomass=c(50,20,10)) %>%
  simulate()
```

set_window	<i>Set window length</i>
------------	--------------------------

Description

Exposure windows are defined as a period of time at the scale of the exposure series. As an example: if an exposure series has an hourly time step, a window length of 24 will consider the exposure within 24 hours intervals for effect calculation. The same applies for the window interval, i.e. the period between considered exposure windows. Set length=-1 to disable moving windows.

Usage

```
set_window(x, length, interval)
```

Arguments

x	vector of EffectScenario objects
length	numeric, length of exposure window to consider for effect calculation, set length=-1 to disable moving windows
interval	numeric, interval between considered exposure windows

Value

modified EffectScenario objects

Examples

```
# calculate the maximum effect for all windows of 10 days length
metsulfuron %>%
  set_window(length=10, interval=1) %>%
  effect()
```

simulate	<i>Simulate an effect scenario</i>
----------	------------------------------------

Description

The supplied EffectScenario is passed on to the ODE solver for numerical integration. Internally, simulate() is split up into several functions dedicated to particular models, e.g. one for GUTS and one for Lemna type models. The package will take care of using the correct function for each model when simulate() is called.

Usage

```
simulate(x, ...)

## S4 method for signature 'EffectScenario'
simulate(x, ...)

## S4 method for signature 'Transferable'
simulate(x, ...)

## S4 method for signature 'ScenarioSequence'
simulate(x, ...)
```

Arguments

```
x          scenario to simulate
...        additional parameters passed on to ODE solver
```

Details

Simulation results are returned as a time-series for each state variable. Some models provide additional properties describing the model state, e.g. the internal concentration of a toxicant within the organism. Refer to the respective [scenario](#) for more information.

Additional arguments to `simulate()` will be passed on to `deSolve::ode()` which enables control of the numerical integration parameters.

Output times and windows:

The minimum and maximum of given time points generally define the simulated period. Function argument `times` overrides settings of the scenario, i.e. time points set in `scenario@times`. However, the simulation can be limited to a subset of time points by enabling a moving exposure window, see [set_window\(\)](#).

Results will be returned for each time point. Precision of the numeric solver may be affected by chosen output times in certain cases. Hence, small deviations in results should be expected if different output times are set. This effect can be mitigated by either defining sufficiently small time step for the solver using argument `hmax` or by decreasing the error tolerances `atol` and `rtol`. These arguments are passed to the solver, see e.g. [deSolve::lsoda\(\)](#) for details.

Optional output variables:

Some models support adding intermediary model variables to the return value of `simulate()`. Analyzing the additional outputs may be helpful to understand model behavior and support finding potential issues in model parameterization.

Optional outputs are enabled by setting the parameter `nout` to a value greater than zero. If `nout` is set to `n`, then the first `n` optional output columns will be returned along the normal simulation result.

Which optional outputs are available depends on the model/scenario at hand. Please refer to the model documentation for details. As an example, the [GUTS-RED-IT](#) model supports adding the external toxicant concentration to the output by setting `nout=1`:

```
minnow_it %>% simulate(nout=1)
```

Numerical precision and stability:

Each model was assigned a default ODE solver which handles most of the occurring inputs well. In most cases, this will be an explicit numerical scheme of the Runge-Kutta family with variable step width. For certain extreme parameters settings, such as very high uptake/permeability of the contaminant or exposure series which represent step functions, the numerical approximation might deteriorate and yield invalid results. In this case try to decrease the allowed max step width by setting the argument `hmax` with various values. Start with `hmax=1` and decrease the value by orders of 10. It is not possible or desirable to reduce `hmax` to extremely small values, as the ODE solver will require more CPU time and simulation will become inefficient.

Oftentimes, it will be computationally more efficient to adapt the solver's error tolerances `atol` and `rtol` than reducing the step width `hmax` to achieve stable numerics. Start by decreasing `deSolve`'s default values by orders of ten until the simulation yields acceptable results, see e.g. [`deSolve::lsoda\(\)`](#) for more information on error tolerances.

As an alternative to adapting solver parameters, it might be worthwhile to try other numerical schemes which can handle stiff ODEs, such as Radau, LSODA, or LSODES. To change solvers, set the `method` argument. To select e.g. the Radau scheme, set `method="radau"`. For LSODA, set `method="lsoda"`. Radau performs better than LSODA in some cases, as the latter method can return biologically nonsensical results without raising an error. See [`deSolve::ode\(\)`](#) for details on available ODE solvers.

Value

A data.frame with the time-series of simulation results

Examples

```
minnow_sd %>% simulate() # tidy syntax
simulate(minnow_sd) # base R syntax

# Set new output times
minnow_sd %>% simulate(times=c(0,4))

# Modify simulated time frame
minnow_sd %>% simulate(times=c(0,10))

# Use an alternative exposure profile than defined in the scenario
minnow_sd %>% set_exposure(data.frame(t=0,c=10), reset_times=FALSE) %>% simulate()

##
## Precision of results

# A large number of output times forces smaller solver time steps
minnow_it %>% simulate(times=seq(0,1,0.001)) %>% tail()

# Defining only two output times allows for larger solver time steps
minnow_it %>% simulate(times=c(0,1))

# The difference between results can be reduced by limiting the solver's
# maximum step length
minnow_it %>% simulate(times=c(0,1), hmax=0.001)
```



```

# The same numerical precision can be achieved by switching to
# the Radau scheme
minnow_it %>% simulate(times=c(0,1), method="radau")

# A very small step length may reduce the difference even further but may
# also exceed the allowed number of steps per output interval. The following
# simulation will be aborted with a solver error:
try(
  minnow_it %>% simulate(times=c(0,1), hmax=0.0001)
)

# However, the solver's max number of allowed steps can be increased:
minnow_it %>% simulate(times=c(0,1), hmax=0.0001, maxsteps=10^5)

```

simulate_batch

Batch simulation using multiple exposure series

Description

[Experimental] A convenience function to simulate a single base scenario with one or more exposure series. This aims at reproducing the setup and results of common effect studies.

Usage

```
simulate_batch(model_base, treatments, param_sample = deprecated())
```

Arguments

model_base	effect scenario object with mean parameters
treatments	treatments exposure levels as data frame (time, conc, trial)
param_sample	<i>deprecated</i> parameter, no longer in use

Details

A scenario contains only one exposure series. However, laboratory experiments commonly examine the effects of multiple exposure levels on a biological system. A batch simulation approach would involve running multiple simulations with varying exposure or treatment conditions. To illustrate, if the objective is to examine the impact of a substance on cell growth, the simulation model could be designed to replicate the cell growth dynamics under varying concentrations of the substance. Each simulation run would represent a specific exposure level, ranging from low to high concentrations of the chemical. To simulate such a laboratory experiment, the `simulate_batch` function can be used. All exposure series are saved in the treatment argument. The first column contains the time, the second column the concentration, and the third column the trial name (exposure level, e.g. 'T1', 'T2', 'T3').

Value

a `data.frame` with simulation results

Examples

```
t1 <- data.frame(time=0:10, conc=0, trial="control") # 1st treatment level
t2 <- data.frame(time=0:10, conc=1, trial="T1")      # 2nd treatment level
treatments <- rbind(t1, t2)

metsulfuron %>%
  simulate_batch(treatments)
```

 solver

Calls ODE solver for a particular model

Description

Please refer to the *Modeling Howto* vignette on how to implement custom models by overloading the solver function.

Usage

```
solver(scenario, times, ...)

## S4 method for signature 'ANY'
solver(scenario, times, ...)

## S4 method for signature 'AlgaeWeber'
solver(
  scenario,
  times,
  approx = c("linear", "constant"),
  f = 1,
  method = "lsoda",
  hmax = 0.1,
  ...
)

## S4 method for signature 'AlgaeTKTD'
solver(
  scenario,
  times,
  approx = c("linear", "constant"),
  f = 1,
  method = "lsoda",
  hmax = 0.1,
  ...
)

## S4 method for signature 'AlgaeSimple'
solver(
```

```
    scenario,
    times,
    approx = c("linear", "constant"),
    f = 1,
    method = "ode45",
    hmax = 0.01,
    ...
)

## S4 method for signature 'DebAbj'
solver(scenario, times, ...)

## S4 method for signature 'DebTox'
solver(
  scenario,
  times,
  approx = c("linear", "constant"),
  f = 1,
  rule = 2,
  method = "ode45",
  ...
)

## S4 method for signature 'DebDaphnia'
solver(scenario, times, ...)

## S4 method for signature 'GutsRedSd'
solver(scenario, times, ...)

## S4 method for signature 'GutsRedIt'
solver(scenario, times, ...)

## S4 method for signature 'LemnaSetac'
solver(scenario, times, ...)

## S4 method for signature 'LemnaSchmitt'
solver(scenario, times, ...)

## S4 method for signature 'MyrioExp'
solver(scenario, times, ...)

## S4 method for signature 'MyrioLog'
solver(scenario, times, ...)
```

Arguments

scenario	scenario object
times	numeric vector of output times, overrides any scenario setting

...	additional parameters passed on to <code>deSolve::ode()</code>
approx	string, interpolation method of exposure series, see <code>deSolve::forcings</code>
f	if approx="constant", a number between 0 and 1 inclusive, see <code>deSolve::forcings</code>
method	string, numerical solver used by <code>deSolve::ode()</code>
hmax	numeric, maximum step length in time, see <code>deSolve::ode()</code>
rule	if approx="constant", a number between 0 and 1 inclusive, see <code>deSolve::forcings</code>

Details

Please note that not all solvers support setting the parameters listed here. In addition, some solvers may set reasonable default values for e.g. maximum step length in time, but not all do. Please check the model documentation for details.

Value

data.frame with simulation results

Methods (by class)

- `solver(ANY)`: Default solver, raises an error
- `solver(AlgaeWeber)`: numerically integrates *Algae_Weber* models
- `solver(AlgaeTKTD)`: numerically integrates *Algae_TKTD* models
- `solver(AlgaeSimple)`: numerically integrates *Algae_Simple* models
- `solver(DebAbj)`: Numerically integrates *DEB_abj* models
- `solver(DebTox)`: Numerically integrates *DEBtox* scenarios
- `solver(DebDaphnia)`: (deprecated) Numerically integrates *DEBtox_Daphnia* scenarios
- `solver(GutsRedSd)`: Numerically integrates *GUTS-RED-SD* models
- `solver(GutsRedIt)`: Numerically integrates *GUTS-RED-IT* models
- `solver(LemnaSetac)`: Numerically integrates *Lemna_SETAC* models
- `solver(LemnaSchmitt)`: Numerically integrates *Lemna_Schmitt* models
- `solver(MyrioExp)`: Numerically integrates *MyrioExp* models
- `solver(MyrioLog)`: Numerically integrates *MyrioLog* models

survival	<i>Survival rate</i>
----------	----------------------

Description

[Deprecated] Derives the survival rate of individuals for *Reduced GUTS* models. Function was replaced by output of `simulate()` and will be removed in a later version.

Usage

```
survival(scenario, ...)
```

Arguments

scenario	an EffectScenario to simulate
...	additional parameters passed on to <code>simulate()</code>

Details

The survival rate describes the survival probability at each time point. The function simulates the *GUTS* scenario and appends a column `survival` to the simulation result. A value of one (1.0) denotes that all individuals survive. A value of zero (0.0) denotes that no individuals survived.

Only available for *Reduced GUTS* models, see [GUTS-RED-models](#). The equations were described by EFSA (2018).

Value

a `data.frame` containing simulation results

References

EFSA PPR Panel (EFSA Panel on Plant Protection Products and their Residues), Ockleford C, Adriaanse P, Berny P, et al., 2018: *Scientific Opinion on the state of the art of Toxicokinetic/Toxicodynamic (TKTD) effect models for regulatory risk assessment of pesticides for aquatic organisms*. EFSA Journal 2018; 16(8):5377, 188 pp. doi:10.2903/j.efsa.2018.5377

See Also

[GUTS-RED-models](#)

Examples

```
# calculate survival rate
minnow_it %>% survival()

# plot survival over time based on a random exposure profile
minnow_sd %>%
  set_exposure(data.frame(t=1:100, c=runif(100)*10)) %>%
```

```
survival() -> df
plot(df$time, df$survival, "l")
```

Transferable

Biomass transfer class

Description

By inheriting from class Transferable, a scenario's behavior can be extended to support transfer and reset of biomass at dedicated points during simulation.

Slots

`transfer.times` numeric, vector of time points at which transfers occur, e.g. `c(7, 10, 14)`
`transfer.interval` numeric, interval length until frond transfer to new medium
`transfer.biomass` numeric, amount of biomass transferred to new medium
`transfer.comp.biomass` character state variable which describes biomass
`transfer.comp.scaled` character vector of state variable which will be scaled 1:1 when biomass is modified, e.g. internal toxicant mass

Biomass transfer

Models supporting biomass transfer can be instructed to move a fixed amount of biomass to a new medium after a period of time. This feature replicates a procedure occurring in e.g. Lemna effect studies and may be necessary to recreate study results.

The biomass transfer feature assumes that always a fixed amount of biomass is transferred. Transfers can occur at any fixed point in time or in regular intervals. During a transfer, the biomass is reset to the transferred amount and additional compartments can be scaled 1:1 accordingly, to e.g. reflect the change in internal toxicant mass when biomass is modified. Transfer settings can be modified using [set_transfer\(\)](#).

Any transfer time point must also be an output time point. If a transfer occurs, simulation results of that time point will report the model state **before** the transfer. Be aware that in order to use transfers at regular intervals, the simulation must start at time point zero.

See Also

[set_transfer\(\)](#)

Other scenario-related: [Scenarios](#)

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